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NANOSIZED DRUG DELIVERY SYSTEMS FOR PLATINUM-BASED ANTICANCER DRUGS

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Abstract. The current review highlights the main advances achieved in utilizing nanosized carriers for optimized drug delivery of cisplatin and other cytotoxic platinum coordination compounds. The treatise is emphasized on exemplary types of carriers (e.g. liposomes, nanoparticles, polymeric micelles, dendrimers, inclusion complexes), their generic properties, advantages, drawbacks, biopharmaceutical and pharmacological aspects as well as the major outcomes of the reported in vitro and in vivo investigations.

Key Words: antineoplastic agents; targeted drug delivery; EPR effect; PEG; liposomes; nanoparticles; polymeric micelles; dendrimers; molecular hosts

Introduction

The platinum coordination compounds are unequivocally among the most important antineoplastic agents used at present for the management of solid tumors [1-3]. The successful commercialization and clinical introduction of the prototype cisplatin in the early 1970's has revolutionized the management of testicular cancer – which is until now among the few examples of successfully curable solid malignancies [3-5]. Furthermore cisplatin is widely used for the treatment of ovarian cancer, head and neck squamous cell carcinoma and has an important palliative role in many other solid tumors [3]. Despite the clinical success of cisplatin, however, it suffers from poor selectivity upon malignant cells and is associated with significant toxic effects upon the kidneys, the peripheral nerves and the auditory system; moreover, cisplatin is the most emetogenic antineoplastic agent [6, 7]. Apart from this unfavorable toxicological profile, there are other significant hurdles limiting the usefulness of cisplatin: the intrinsic unresponsiveness of some common neoplastic diseases e.g. colon adenocarcinoma, the development of acquired resistance in initially responsive tumors in the course of treatment [1, 8]. Thus the clinical efficacy of cisplatin, together with its major limitations have fuelled intensive research efforts focused upon the development of analogues with better tolerability, broader antineoplastic spectrum and/or superior activity in comparison to the prototype. To meet these objectives thousands of platinum coordination compounds have been synthesized and evaluated, which up to now resulted in the successful development and commercialization of five clinically utilized cisplatin analogues (Depicted in Fig. 1) [1, 4, 5]. Among these carboplatin and oxaliplatin have widespread clinical use [1], whereas the remaining drugs are available only in particular countries - nedaplatin in Japan [9], lobaplatin in China [2, 10] and heptaplatin in South Korea [11-14]. Unfortunately none of the successors of cisplatin could be considered superior to the prototype in terms of lower toxicity, superior clinical efficacy and bypassing resistance mechanisms; for instance nedaplatin and carboplatin are totally cross-resistant with cisplatin and the significant amelioration of nephrotoxicity characteristic for these drugs is achieved at the expense of reduced cytotoxicity against malignant cells [2]. Oxaliplatin, although showing low degree of cross-resistance with cisplatin does not affect a vastly different spectrum of tumors and has rather limited clinical utility so far [1]. Furthermore despite the low nephrotoxicity of the novel platinum drugs they display modified rather than reduced toxicological potential compared to cisplatin, whereby the dose-limiting toxicity is switched to myelosuppression in carboplatin, nedaplatin and lobaplatin and to severe peripheral neuropathy in oxaliplatin [1, 2, 9].

The failure of existing platinum drugs to overwhelm the limitations of cisplatin as an anticancer drug, especially regarding the issues of resistance could be greatly ascribed to their structural resemblance to the prototype and hence their closely-related biochemical and pharmacological properties. More recently the advances of bioinorganic chemistry and the unraveling of the mode of action of cisplatin and the resistance mechanisms gave rise to more elaborate and rational approaches for designing promising anticancer platinum complexes [4, 5].

Fig.1 Chemical structures of the most important clinically used platinum-based anticancer drugs

Apart from the lengthily and expensive design and elaboration of novel analogues one of the most attractive alternative strategies to overcome the limitations of cytotoxic drugs, including platinum metallopharmaceuticals is their formulation into nanopharmaceutical platforms, i.e. nano-scale carriers, such as liposomes [15-17], polymeric nanoparticles (nanospheres, nanocapsules, polymeric micelles, multiarm core-shell co-polymers, protein or polysaccharide conjugates etc.) [18-20], and more recently into nano-containers based on host-guest interactions [21-26]. Due to their unique properties the nanopharmaceuticals offer significant advantages over classical parenteral formulations of anticancer drugs and have been well demonstrated to decrease drug binding to non-pharmacological targets, to favorably alter the systemic and intratumoral trafficking of encapsulated agents and to greatly ameliorate the debilitating dose-limiting toxicities, associated with this class of antineoplastic drugs [27, 28].

To a great extent this is due to the fact that drugs are encapsulated within nanocontainers with a controlled microenvironment, whereby the drug is protected from side interactions with body tissue components, xenobiotic efflux transporters and biotransformation systems. Thus the pharmacokinetic and tissue distribution of a drug encapsulated in a nanoplatform are no more dependent on its intrinsic properties, but are governed by the tissue disposition and elimination patterns of the carrier [29, 30]. Moreover, additional benefits of nanoparticulate systems include sustained or trigerrable release kinetics, increased bioavailability at the respective targets sites with concomitant increased efficacy, reduction of the nominal dosage required and amelioration of the severity and incidence of adverse reactions [29-31].

This review is focused on representative examples of nanopharmaceutical platforms for platinum coordination compounds with special emphasize on liposomes, globular architecture polymeric nanoparticles (micelles, dendrimers and stars) and macrocyclic molecular hosts.

Strategies for passive of active tumor targeting of platinum drugs

It is well known that the growth of solid malignant tumors is dependent on a process of *de novo* formation of blood vessels known as angiogenesis [32, 33]. The newly formed vasculature of tumors however is leaky relative to the vessels in normal tissues which makes solid tumors hyperpermeable towards colloid-sized carriers, e.g. liposomes and polymer nanoparticles [16, 34, 35]. The compromised barrier function of the vasculature, together with the inadequate lymphatic drainage of tumors conditions the accelerated accumulation of blood-borne nanoparticles, i.e. the 'enhanced permeability and retention effect' (EPR effect) has been the central paradigm that has fuelled the development of antineoplastic nanopharmaceuticals during the last three decades [32, 34-38].

One of the hallmark challenges associated with nanocarriers is that these have to circulate long enough in order to attain enough accumulation at tumor lesions *via* the EPR [30, 39, 40]. Due to the colloidal size of nanocarriers these are recognized and phagocytized by the cells of the mononuclear phagocyte system (MPS) (previously designated as reticuloendothelial system) which leads to disappointingly short

circulation half-lives [16, 41-43]. The most important approach towards bypassing MPS sequestration has been the incorporation of PEG residues on the surface of polymer particles or liposomes [39, 44-47]. PEGylation creates a hydrophilic repulsive barrier around nanocarriers which increases their colloidal stability, hinders interactions with serum components and opsonins, and eventually prevents recognition by the MPS cells [39]. This imparts MPS-avoidance or "stealth" properties to the delivery device, increasing its systemic circulation time significantly [27, 39, 46, 48]. Moreover, PEGylation of macromolecular carriers has been well documented to favorably decrease their immunogenicity [49], although some of the adverse effects associated with stealth liposomes, have been attributed to immune responses.

While the EPR-driven passively targeted systems have been extensively explored and dominate the commercialized and clinically tested nanopharmaceutical medicinal products these are by no means ideal substitutes for the conventional dosage forms of anticancer drugs. Albeit EPR effect results in selective delivery progressively over time, it has been shown that PEGylation while beneficial for the circulation time is detrimental for the cellular uptake of nanocarriers [16, 40]. Thus although anticancer drugs are selectively accumulated inside tumors they are retained within the carrier, which could result in subtherapeutic bioavailability inside cells, reduced efficacy and emergence of drug-resistance [16]. Hence there is a need for attaining augmented intracellular and localized, on-demand drug release in order to beneficially modify the efficacy of nanopharmaceuticals [27, 28]. On these grounds state-of-the art research has been shifted from EPR-driven systems towards more sophisticated nano-vehicles for actively targeted drug delivery of antineoplastic agents [27-29, 50]. This is achieved by surface decoration of the nanoplatforms with homing ligands ensuring site specific delivery or by use of "smart" technologies attaining triggered, on-demand release of encapsulated cargo in response to environmental stimuli or remote triggers, e.g. ultrasonication, hyperthermia, and magnetic fields [18, 20, 27, 28, 51].

Surface decoration of nanocarriers with ligands able to bind with high affinity membrane receptors over-expressed in tumor cells is among the most widely explored approaches towards tumor targeting [18, 27, 28]. If such modification is concomitant with long circulating properties and ability to passively accumulate inside solid tumors *via* the EPR effect, the active targeting is expected to further increase the

specific interactions with cancer cells. Moreover, if the targeting ligand is binding to internalizing surface epitopes of cancer cells is could also aid to increased intracellular uptake of the carrier and its cargo *via* receptor-mediated endocytosis [18].

If the receptor is highly specific for the tumor cell usually the ligands are coupled to the PEG coating of the nano-carrier (i.e. pendant type ligands) [40, 44]. In a dissimilar fashion if the pendant-type ligand is expected to bind to receptors outside tumors, or to evoke immune recognition, both leading to decreased tumor accumulation of the carrier, the homing moiety could be grafted on the surface of the nanoparticle and hence its interactions with the receptor would be hampered by the protective PEG coating [44]. The most widely used homing moieties to ensure active targeting of nanoparticles and liposomes are monoclonal antibodies (mABs) [45]; small molecule cofactors and vitamins such as folate [52], riboflavin [53], biotin [54-56]; hormones and hormonal analogues [57-60]; transferrin [61, 62], carbohydrates [63, 64], ligands for adhesion molecule receptors [65-68], and aptamers [69-73], among others. The mechanistic rationale for active targeting is briefly outlined in the following section, whereas recent examples of targeted liposomal or polymeric nanocarriers are summarized in the respective sections of the review.

One of the feasible approaches for triggered drug release at the tumor site explores the lower pH of the intratumoral and other microenvironments, relative to the dominant physiological pH of 7.4 characteristic for the majority of body fluids [74-78]. Such mildly acidic conditions exist in tumor and inflammatory tissues (pH 6.8) and in endosomes (pH 5-6) [78]. It is well established that one of the most important mechanisms for cellular uptake of nanopharmaceuticals is endocytosis. Once the particle is internalized inside cells by endocytosis, the endocytic vesicles eventually evolve to late endosomes and then to lysosomes, whereby the acidity (pH 5.0) is significantly higher as compared to the general physiological condition (pH 7.4) . Unfortunately, if the drug is polar and chemically unstable (e.g. the active diaqua-metabolites of platinum agents) it could be retained inside these intracellular compartments, unable to reach its ultimate pharmacological targets, and eventually degraded by via to hydrolysis enzymatic cleavage. By achieving endosomal escape of encapsulated cargo pH-responsive vehicles increase the cytosolic bioavailability of free drug and hence are expected to augment its cytotoxic effects [79, 80].

Thus pH-sensitive systems, depending on their architecture, properties and acidity triggered destabilization kinetics have the advantage of site-specific drug release either within the intratumoral microenvironment or within the cytosole of tumor cells [18, 79-82]. The acidity-responsive nano-carriers that have been most widely explored are pH-sensitive liposomes and polymeric micelles. The pH-triggered release from liposomes is usually attained *via* destabilization and increased fusogenicity of the liposome membrane under acidic conditions [77, 80, 83].

Liposomal platinum drugs

Liposomes are spherical vesicles comprising either a single phospholipid bilayer or alternating tightly packed aqueous compartments and lipid bilayers which enclose a central aqueous reservoir. Liposomes are nano- to micro-sized vesicles (50–5000 nm) and according to mean diameter and lamellarity fall into two categories, namely multilamellar vesicles (MLVs) with a size of 500–5000 nm and unilamellar vesicles (ULVs) with a size range of 50–250 nm. ULVs are further separated into small unilamellar vesicles (SUVs) sized 50-100 nm and large unilamellar vesicles (LUVs) with diameters exceeding 100 nm [84].

Liposomes are composed of naturally derived or synthetic phospholipids which conditions their excellent biocompatibility, biodegradability, non-immunogenicity and generally low toxicological and safety pharmacological potential [29, 30, 50, 85]. Due to the abundance of lipid and aqueous compartments liposomes are capable of accommodating both polar and non-polar compounds based on their solubility and partitioning characteristics [84, 86]. Lipophilic agents are typically encapsulated within the lipid bilayer of liposomal membranes, whereas polar agents are confined to the aqueous central cavity. Due to the possibility of spontaneous leakage and hydrolysis in the aqueous phase the liposomes the entrapped polar compounds face more challenges in terms of chemical and biopharmaceutical stability [84].

Several milestone advances in liposome technology have been made to meet some of the pharmaceutical and pharmacokinetics challenges associated with the drug loading efficiency, biodistribution and targeting potential of liposomes that allowed the evolution of these systems from membrane models to elaborated drug delivery platforms. These involve the advent of: techniques for size reduction and homogeneity of liposomal populations, the remote drug loading maneuvers allowing significant entrapment

efficiency for weakly acidic or basic drugs [84], the realization of the steric stabilization concept [86], and the development of on-demand triggerable or fusogenic liposomes [82, 85, 87]. Thus over the last three decades the liposomes platforms have evolved from plain or conventional liposomes to long circulating "stealth" liposomes and ultimately to targeted and stimuli-responsive liposomes [44]. Moreover, the possibilities for complex surface modification, conjugation of permeation enhancing or homing moieties, incorporation of pH-responsive components etc. condition the possibilities for designing sophisticated drug delivery platforms with hybrid tumor- targeted and triggerable release properties [82].

Conventional Liposomes

The non-modified conventional liposomes are mainly composed of natural or synthetic phospholipids such as 1,2-distearoryl-sn-glycero-3-phosphatidyl choline (DSPC), 1,2-dipalmitoyl-sn-glycero-3-phosphatidyl choline (DPPC), egg phosphatidylcholine, etc., and cholesterol, without polymer coating or other steric stabilizing moieties [15, 16, 84]. Despite their favorable biphasic nature and low toxicological potential these are associated with inherited obstacles including lack of specific targeting and propensity to promptly accumulate in MPS organs with consequent fast disappearance from the circulation and disappointingly low bioavailability except in the liver and spleen [16, 40, 84]. This behavior is due to side interactions between liposomes and serum components, such as LDL and HDL leading to membrane destabilization, favoring eventual opsonization by a2-macroglobulin, fibronectin, blood clotting factors, complement components, among others [88]. Thereafter the opsonin-tagged liposomes are recognized and phagocyted by macrophages, Kuppfer cells and other MPS cells (Fig. 2) [40, 88].

To address these issues different approaches have evolved including manipulations of particle size, surface charge, phospholipid bilayer content *etc.* [16, 86, 88]. Thus addition of cholesterol has proved to increase the packing density and integrity of bilayers, hampering serum protein interactions and opsonization and increasing the ability of liposomes to retain their aqueous cargo within the circulation [88]. Moreover, it has been shown that reduction of the particle size is a highly successful strategy to evolve MPS sequestration as multilamellar liposomes with sizes ranging 500–5000 nm are far more promptly eliminated from the circulation as compared to the more recently developed large unilamellar liposomes or

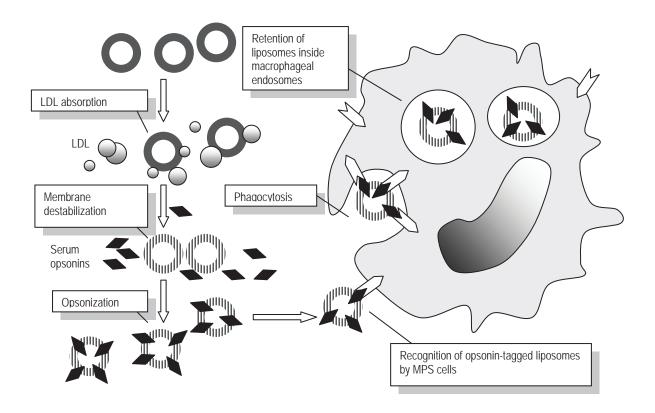


Fig.2 Opsonization, recognition and phagocytosis of conventional liposomes by the cells of the mononuclear phagocyte system (MPS)

the low nano-scale sized small unilamellar liposomes [16]. Despite the decreased propensity to accumulate in MPS organs the small unilamellar liposomes share the serious disadvantage of having relatively lower aqueous volumes. Hence they are characterized with generally lower drug entrapment capacity. In general, elimination or reduction of the surface charge is another favorable approach as both anionic and cationic liposomes appear to have shorter half-lives and increased intrinsic toxicity [16, 88].

Intensive research has been focused at liposomal delivery systems for platinum agents [5, 8, 89]. A representative advance in this field is cis-(bisneodecanoato)-trans-R,R-1,2-cyclohexanediamine platinum(II) (NDDP), characterized *via* high liposomal encapsulation efficacy which is currently developed by Aronex Pharm. Inc. Aroplatin® demonstrated promising activity in diverse experimental tumors and received FDA-orphan drug designation for the treatment of malignant mesothelioma [5, 8, 89]. This formulation has been subject to clinical trials with patients with advanced solid tumors or B-cell lymphoma, and is currently undergoing phase II clinical trials in colorectal carcinoma [27].

Another water insoluble agent 2-(4-(tetrahy-dro-2H-pyran-2-yloxy)-undecyl)-propane-1,3-diam-

minedichloroplatinum(II) (THP-C11) was incorporated in LUVs. The liposomal formulation (LipoTHP-C11) showed excellent stability at 4°C for more than two months. It proved to exert cytotoxic effects in a panel of cell lines: H12.1, 1411HP, 518A2, A549, HT-29, MCF-7 and SW1736, with concomitant lower activity towards normal human fibroblasts [90].

"Stealth" Liposomes

Owing to the circumvention of the most important problems peculiar with conventional liposomes, namely the short circulation half life and the prompt sequestration in MPS organs the long-circulating or "stealth" liposomes have earned their place as dominant drug delivery platforms for anticancer drugs to be subject of both preclinical and clinical studies [16, 40, 91, 92]. Stealth liposome strategy is based on the process of steric stabilization, i.e. creating a repulsive polar coating around vesicles by grafting liposomal membranes with lipid-anchored hydrophilic polymers [39]. To meet this objective a number of different natural and synthetic polymers have been employed as steric stabilizing agents. These include, but are not limited to polyethylene glycol (PEG), chitosan, polysialic acids, polyvinyl alcohol (PVA), poloxamers, poly(acrylamide), poly(vinyl

pyrrolidon), poly(acryloyl morpholine), poly(2methyl-2-oxazoline), poly((2-ethyl-2-oxazoline), poly(vinyl alcohol), hydroxypropylmethylcellulose, etc. [16, 40, 88]. In line with its biocompatibility, pharmaco-toxicological inertia, acceptably low immunogenicity as well as because of its extensively validated efficacy as steric stabilizing agent PEG has remained the golden standard for engineering long circulating liposomal nanoplatforms [27, 39]. PEGylated liposomes are most often prepared by introduction of distearoylphosphatidylethanolamine-PEG-2000 (DSPE-PEG-2000) (Fig. 3) which hampers their interactions with HDL and serum opsonins, and conversely decreases the capacity of MPS cells to recognize and phagocyte liposomes (Fig. 3), leading ultimately to significantly increased plasma half-lives [93].

The efficacy of the stealth liposome concept for attaining optimized tumor-site bioavailability and its comprehensive experimental and clinical justification have fuelled immense and enduring efforts for elaboration of PEGylated-liposomal antineoplastic agents, such as platinum drugs [94-99]. Exemplary stealth liposomal formulations evaluated in clinical trials are summarized in Table 1, and reviewed elsewhere [100, 101].

An important example of clinically validated liposomal formulations is SPI-077 – a stealth-liposomal cisplatin. This platform is featured by a high encapsulation efficiency, favorably tailored biodisposition

and pharmacokinetics and conversely ameliorated toxicity relative to the free drug. Despite these beneficial characteristics however SPI-077 has failed to demonstrate prominent efficacy advantages compared to cisplatin presumably due to the very slow release of encapsulated cargo [8].

A more recent liposomal drug delivery system of cisplatin under clinical development is LipoplatinTM, elaborated by Regulon Inc. [102, 104]. This stealth liposomal system has shown significant advantages compared to free cisplatin in both preclinical and clinical settings, in terms of improved intratumoral accumulation, avoidance of the hallmark toxicity, concomitant with significant antineoplastic efficacy equal or even superior to that of the non-encapsulated drug [102, 104, 106]. LipoplatinTM is currently undergoing several phase II and phase III trials in combinations with other antineoplastic agents, such as gemcitabine, 5-fluorouracil, and vinorelbine [2, 102-104, 106-109]. The same company has developed a stealth liposomal formulation of oxaliplatin (LipoxalTM) whose preclinical development has shown promising effects in resistant tumor models [94] and potent radiosensitizing activity in F98 glioma [95, 96]. LipoxalTM is currently subject to clinical evaluation. A Phase I trial in patients with progressive and recurrent gastrointestinal cancers has shown that the product is well-tolerated and greatly ameliorates the non-neurological toxicities of oxaliplatin [105].

Table 1. Stealth liposomal or non-PEG polymer coated liposomal formulations of platinum anticancer drugs in clinical trials

API	Name	Formulation	Indications
Cisplatin	SPI-077	Stealth liposomes (i.v.); HSPC, CHOL and mPEG-DSPE	Head and neck cancer, lung cancer (Phase I-II) [8]
Cisplatin	Lipoplatin TM	Stealth liposomes (i.v.); SPC, DPPG, CHOL , mPEG-DSPE	Several cancer types; (Phase II-III) [102-104]
Oxaliplatin	Lipoxal TM	Stealth liposomes (i.v.)	A completed Phase I trial in advanced GIT cancer [105]

Abbreviations: API: Active pharmaceutical ingredient; HSPC: Hydrogenated Soy Phosphatidylcholine; DPPG: Dipalmitoylphosphatidylglycerol; mPEG-DSPE-poly(ethyleneglycol)-distearoylphosphatidylethanolamine.

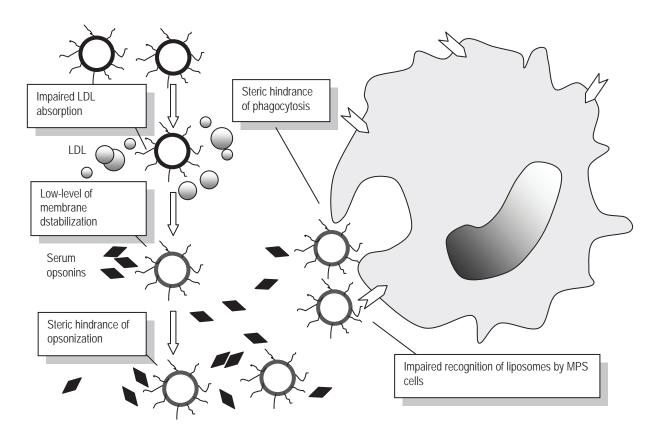


Fig.3 Impact of steric stabilization against MPS recognition and sequestration of "stealth" liposomes. The densely packed and PEG-grafted bilayers resist LDL interactions with subsequent low level of opsonization and hampered recognition and phagocytosis by mononuclear phagocytes (See the text for details)

Targeted Liposomes

The targeted liposome based drug delivery concept was fuelled by the established incapacity of non-decorated stealth liposomes to avoid exposure of non-malignant tissues and to eliminate the hallmark dose-limiting toxicities of cytoreductive chemotherapy [27]. These site-specific targeted liposomes are decorated with different types of homing moieties to increase the rate of liposomal drug accumulation in the ultimately targeted tissues/cells *via* interactions with cancer cell receptors/antigens (Fig. 4) [15, 85, 92].

Peptide, incl. peptide hormones and proteins have been also widely explored as targeting ligands for liposomal antineoplastic drugs. Among these special attention has been paid to transferrin-modified vesicles [61, 62, 143-148]. As with other targeting ligands, Tf could be conjugated or anchored directly to bilayer membranes of plain liposomes [16, 61, 62, 148, 149], or *via* the PEG-coating of stealth liposomes, a strategy which is has been increasingly employed in recent investigations as it allows to combine the plasma longevity of stealth liposomes with

the enhanced cellular accumulation *via* Tf-receptor mediated endocytosis [44].

A recent contribution evaluated the cytotoxicity and cellular accumulation of Tf-modified cisplatin-loaded PEGylated liposomes in chemosensitive and cisplatin-resistant A2780 cells. Free cisplatin was 4 times less efficiently accumulated inside resistance cells, whereas the uptake of liposomal drug was comparable in both cell lines. Albeit the cytotoxicity of liposomal drug vs. free cisplatin was somewhat lower in the sensitive cell line, but was significantly higher in the resistant variant, in corroboration to the accumulation kinetics data [151].

Another paper reported cisplatin-loaded liposomes targeted at Tie2 - a receptor tyrosine kinase that plays important roles in vascular angiogenesis, and is highly expressed in vascular endothelial cells and a number of cancer cells. The delivery platform employed a novel peptide ligand PH1 peptide (TMG-FTAPRFPHY) selected by phage display library screening combined with surface plasmon resonance binding assays. The homing moiety was covalently conjugated to the distal end of DSPE-PEG(2000)-

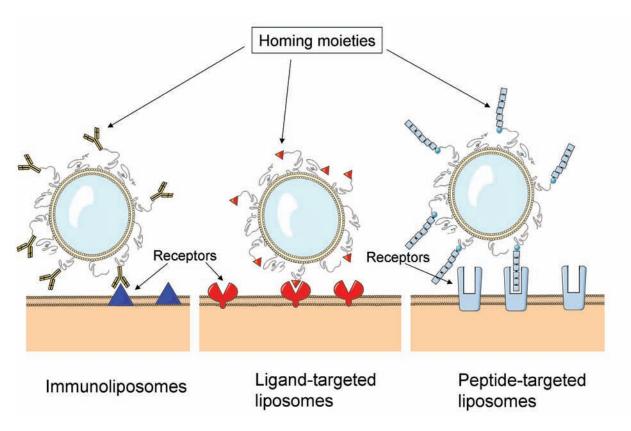


Fig.4 Schematic representation of targeted stealth liposomes

Maleimide lipid and grafted onto liposome membranes. These PH1-PEG-liposomes containing the anticancer drug cisplatin were showed to bind tightly to Tie2 positive cells, mediate active endocytosis of the drug containing liposomes, and result in much higher cell specific cytoxicities than mPEG coated liposomes [153].

pH-Responsive Liposomes

One of the hallmark issues associated with conventional liposomes is their propensity following cellular internalization to accumulate in certain subcellular compartments, mainly lysosomes, where the encapsulated drug is retained or even degraded, thus limiting its availability at the cytosolic target site (Fig. 5). This is of paramount importance in anticancer drug delivery as the majority of antineoplastic agents interact with pharmacological targets located inside cells or within the nucleus (e.g. genomic DNA) [16, 88]. An attractive and vastly explored approach to avoid lysosomal sequestration and degradation of entrapped materials is the use of pH-sensitive liposomes exhibiting considerable fusogenic activity at low pH and capacity to evade endosomal sequestration (Fig. 5) [17, 44, 80, 82, 154].

The typical pH-triggerable systems are composed from dioleoylphosphatidylethanolamine (DOPE) with cholesteryl hemisuccinate (CHEMs) or other acidic amphiphiles acting as bilayer stabilizers at neutral pH. DOPE comprises a compound with minimally hydrated and hence small headgroup, occupying relatively smaller volume vs. the bulky hydrocarbon chains [84]. This imparts a cone shape to the molecule, which is detrimental for bilayer assembly and supports the formation of inverted hexagonal phase micelles. Introduction of an acidic amphiphile which is negatively charged at physiological pH among DOPE molecules allows the formation of bilayer structures, and facilitates the construction of liposomes, stable at physiologically relevant pH and temperature [80, 84, 155]. In acidic pH the protonation of the carboxylic groups of the amphiphiles, reduces their stabilizing effect and this leads to liposomal membrane destabilization, since under these conditions PE molecules revert into their inverted hexagonal phase [80, 155]. More recently new classes of pH-sensitive systems based on fusogenic peptides [147], or surface modification of liposomes with pH-sensitive polymers [138, 154] have emerged, extensively discussed elsewhere [81, 155].

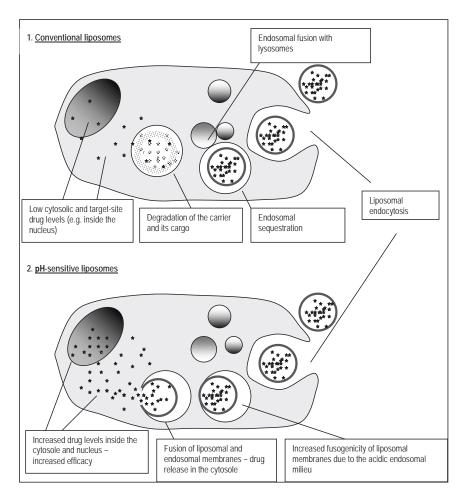


Fig.5 Comparative schematic representation of the fate and trafficking of conventional liposomes, which are retained within the endosomal compartments (above) and pH-sensitive liposomes which become fusogenic inside endosomes and deliver their cargo in the cytosole (below)

An exemplary novel DOPE:CHEMs system is a DSPE-PEG2000-modified formulation of cisplatin, developed as a sterically stabilized pH-responsive platform for intraperitoneal application [97] and optimized in terms of technological and formulation variables to obtain most favorable protocol for a reproducible and economically feasible large scale production [156]. These liposomes proved to be serum stable and to selectively accumulate and induce cytotoxicity in cisplatin-resistant small-cell lung carcinoma cell line (GLC4/CDDP), characterized with reduced drug uptake [97]. Specialized toxicological surveys of this formulation showed pronounced amelioration of the lethal toxicity (with significantly higher LD₅₀ values compared to free drug), and elimination of cisplatin-related myelosuppression and hallmark nephrotoxicity following intraperitoneal [157] or venous application [158] in mice. The long circulating pH-sensitive liposomes of cisplatin

proved to exhibit potent tumor-inhibiting properties following intraperitoneal application against Erlich ascites tumor with more marked increase in life span as compared to the free-cisplatin treated group. Moreover, in this therapeutic intervention the liposomal formulation demonstrated favorable safety characteristics vs. non-encapsulated cisplatin, in corroboration to the toxicological data [159].

Enzymatically Triggerable Liposomes

A sophisticated liposomal drug delivery system based on enzymatically assisted pro-drug activation and carrier destabilization is the LiPlasome® platform [85, 87, 164-167]. This elaborate strategy for triggered release of liposomal cisplatin or other antineoplastic agents is based upon the significant up-regulation of the secretory phospholipase A2 (sPLA2), peculiar for a variety of certain solid tumors [87, 164, 167-171]. To exploit this feature LiPlasome Pharma

A/S has developed liposomal drug delivery system (LiPlasomes®) composed of ether lipid prodrugs which are preferentially degraded within tumors, over-expressing sPLA2 [85, 87, 164-166, 168, 170]. The degradation of the liposomes is consistent with release of encapsulated drugs, and *de novo* formation of cytotoxic ether lipids and membrane disrupting fatty acid residues. The latter further destabilize the membranes of target cancer cells and facilitate the drug accumulation therein [30, 166-168]. The most advanced products of the company are PLA2-triggerable liposomal cisplatin (LiPlaCis®) which is subject to Phase I trials and oxaliplatin (LiPloxa®), currently under preclinical toxicological evaluation [172].

Sonosensitive Liposomes

Another feasible trigger for remote guided release of drugs from liposomes is ultrasound exposure. Ultrasonication has been documented to increase drug release from conventional and stealth liposomes with different phospholipid content [82, 175, 176]. Schroeder et al. have explored the possibilities for ultrasound assisted release of cisplatin from sterically stabilized liposomes. A pharmacokinetic study in Balb/C mice transplanted with C26 colon adenocarcinoma tumors in the footpad showed that the use of low-frequency ultrasound led to almost 70% of liposomal cisplatin release in contrast to 53% when there was no sonication [176]. These findings are especially intriguing having into consideration the fact that the unsatisfactory clinical performance of liposomal cisplatin (SPI-077) is at least partly due to the slow release of the drug from the sterically stabilized liposomes [8].

Polymer-based drug delivery systems

Polymeric micelles and macromolecular pro-drugs

Polymer-drug conjugates are water-soluble tailor-made structures designed to modulate the pharma-cokinetic properties of the therapeutic agent [19, 32, 37, 177]. On the one hand, particulates with a larger size than glomerular excretion threshold value (42–50 kDa for water-soluble polymers) may provide a prolonged blood circulation of the conjugated drug. On the other hand, polymeric carriers having a size smaller than 200 nm and a hydrophilic and biocompatible surface may avoid the recognition by the reticuloendothelial system [27, 28, 32, 35, 37, 38, 178]. Besides optimization of the rate and duration of drug delivery the conjugation strategy can provide drug targeting to specific cells or tissues and control of the

release of highly toxic drugs as an effective way to minimize the adverse side effects. In addition, the hydrophilic polymer carrier can impart favorable physicochemical properties, e.g. increasing the solubility of lipophilic drugs or the stability of labile agents from chemical or proteolytic degradation [27, 28].

It is well-appreciated that cisplatin and its analogues react with a variety of nitrogen- and sulphurcontaining biomolecules by ligand exchange reactions [3, 179]. In blood a high fraction of cisplatin is bound to plasma proteins, including albumin, transferrin and g-globulins that reduce its therapeutic concentration. The ligand exchange kinetics of platinum compounds is largely determined by the nature of the leaving groups. Carboxylate groups possess low nucleophilicity and therefore they are able to undergo the reverse exchange reaction with chloride ions to regenerate cisplatin at physiological salt concentrations [4, 5].

The property of carboxylate ligand as a good leaving group has been exploited to design cisplatin delivery systems based on carboxylate-containing polymers. Polymer-drug complex micelles were spontaneously formed on mixing of cisplatin with PEO-poly(aspartic acid) or PEO-poly(glutamic acid) block copolymers in an aqueous solution [180-184]. The cisplatin-incorporated micelles were extremely stable in distilled water whereas in physiological saline the micelles showed dissociation into unimers, accompanied with sustained platinum (II) complexes release. The micelles formed from PEG-bpoly(aspartic acid) underwent fast structural decay (~30 h) that caused liver and spleen accumulation and comparable antitumor activity to free cisplatin despite restrained nephrotoxicity. The time scale of decaying of the micelles was prolonged to 50 h when PEO-b-poly(glutamic acid) copolymers were used for cisplatin conjugation which improved the selectivity and efficiency in tumor targeting [184].

The metal ligand coordination was also utilized to incorporate the drug into the cross-linked micelles with ionic poly(methacrylic acid) cores and a hydrophilic shell of PEO chains [185]. The size of the loaded micelles was about 150 nm and the drug content was determined to be 22%(w/w). Cisplatin was encapsulated in nanoparticles formed by hydrophobically modified chitosan [186] or thermosensitive polymer carriers [187, 188].

In contrast to the particulate carriers such as micelles, water-soluble polymers allow drug molecules to interact with a single macromolecule rather than a large particle. The polymer carriers take advantage

of EPR effect without accumulating into the liver and spleen. However, linear polymers have limited drug payload capacity. For instance, binding of cisplatin to homopolymers and alternating copolymers bearing carboxylate moieties often results in the formation of poorly soluble cisplatin-polymer conjugates when the molar ratio of cisplatin to carboxylate residues in the polymer exceeded 0.2 [189].

Dendrimers

Dendrimers comprise a class of globular, highly branched, synthetic macromolecules with tunable size and architecture [52, 190, 191]. They encompass multiple layers with large number of chemically active surface groups, also known as generations, which emanate out of an initiator core, denoted as generation zero (G_0) . Typically the size of dendrimer particles ranges 1–15 nm and these are characterized by significant homogeneity in terms of size distribution and morphology [191]. Dendrimers have many attractive properties which make them advantageous drug carriers as compared to both linear and hyperbranched polymer-based systems [191-193]. The unique highly regular branching architecture and the multiple arms of dendrimers provide a large multivalent backbone whereby anticancer drugs, targeting moieties or solubilizing groups can be feasibly attached through covalent conjugation or electrostatic adsorption. Moreover, drugs could be also loaded within the cavities of the core regions, either covalently or via hydrophobic, hydrogen, or van der waals bonding. In addition, the low level of polydispersity of these macromolecules is a prerequisite for reproducible pharmacokinetic and biodistribution behavior which is of paramount importance for their actual applicability as drug delivery systems [191, 194].

The research on dendrimer-drug deliver systems is focused on biodegradable backbones, e.g. the polyaryl ether dendrimers, polyester dendrimers based on 2,2-bis(hydroxymethyl)propionic acid, glyceryl-succinate polyester dendrimers, and especially polyamidoamine (PAMAM) dendrimers [27, 52, 190, 191]. The latter have been exceptionally widely studied as drug delivery platforms in line with their excellent biocompatibility, water solubility, and abundance of large number of active functions suitable for coupling of chemotherapeutic agents and targeting ligands [52, 190, 191].

Considering the ubiquitous abundance of biotin as micronutrient especially in rapidly proliferating cells such as cancer cells Yellepedi *et al.* designed biotinylated PAMAM dendrimers as a targeted carrier for

antineoplastic agents, including platinum drugs. The effect of generation and the mechanism of cellular uptake of biotin-PAMAM-G₄ in ovarian cancer (OV-CAR-3) and human embryonic kidney (HEK 293T) cells was determined by fluorescent microscopy and flow cytometry. The cellular uptake of biotin-PAMAM was significantly higher in the cancer cell line, as compared to the non-malignant HEK293T cells. Mechanistic studies demonstrated that the cellular uptake of biotinylated-PAMAM was mediated by biotin receptor-mediated endocytosis and chargemediated adsorptive endocytosis. The cytotoxicity of biotinylated-PAMAM-G₄ in the HEK 293T cells was comparable to that of the parent PAMAM dendrimers [54]. To further address the applicability of the show potential as nanocarriers in targeted drug delivery the same group developed cisplatin-loaded biotinylated PAMAM dendrimers. The systems were investigated for encapsulation efficiency, in vitro cytotoxic activity and cellular accumulation of cisplatin in a panel of chemosensitive (OVCAR-3, SKOV-3, A2780) and one cisplatin-resistant (A2780/CP70) ovarian cancer cell lines. The PAMAM dendrimers displayed relatively low encapsulation efficiencies of cisplatin ranging ca. 5-21%. The dendrimer loading however significantly augmented the cytotoxic effects of the drug as evidenced by the significantly lower values of the IC₅₀ values thereof vs. those of the free drug. The cytotoxicity data were corroborated by an in vitro accumulation assay which showed that the PAMAM-G₄ NH₂ dendrimer complexes of cisplatin display far more efficient, approximately ten fold higher uptake in both A2780 and A2780/CP70 cells as compared to the free drug. These finding point out for the feasibility of biotinylated PAMAM dendrimers as potential targeted nanoplatforms of cisplatin in ovarian cancer [55].

Multi-arm Star-Like Polymers

In spite of the undisputable advantageous characteristics of dendrimers, e.g. their well-defined and homogenous size distribution patterns controlled branching architecture and drug loading feasibility their synthesis is time consuming and tedious stepwise procedure. Thus an emerging alternative are the core—shell type star polymers bearing hyperbranched cores and multi-arm shell of linear polymers bearing active end functionalities [195, 196]. These new macromolecules based upon various branched core architectures exhibit "unimolecular micelle" behavior in water solution, whereby the covalently linked interior and shell domains remain stable independently

of concentration, abundance of interactive solutes and temperature [197-202]. The structural stability and multifunctionality of the stars conditions the significant scope for their elaboration as drug delivery systems, since they provide opportunities for either chemical/ electrostatic immobilization, or physical encapsulation of anticancer drugs [197, 202].

Another recent report has presented the formulation and evaluation of a core–shell type star polymer with a branched hydrophobic polystyrene interior and covalently attached poly(acrylic acid) arms, as a drug delivery system for cisplatin. This architecture proved to afford several advantages as cisplatin carrier such as high density of carboxylate functions that are able to reversibly immobilize the drug, exceptionally high drug payload, stability in aqueous milieu upon storage and sustained release of the agent under physiological conditions.

The system displayed prominent capability for intracellular uptake and exhibited concentration and time- dependent cytotoxicity in a panel of human tumor cell lines [203]. This cisplatin -formulation has been further developed to a reversibly PEGylated nanocarrier. The design strategy was based on functionalization of the polyacrylate arms *via* a PEGylated cisplatin analog, allowing for detachment of the coating following hydrolysis in biological milieu. The formation of PEG shell resulted in higher drug payload and improved drug release profile of the nano-conjugates. The *in vitro* bioassay confirmed that the PEGylated conjugates exhibited higher cytotoxicity compared to the non-PEGylated cisplatin -loaded stars [204].

Molecular hosts as drug delivery systems for platinum metallodrugs

The supramolecular interactions of macrocyclic hosts with different types of small guest-molecules leading to formation of stoichiometric inclusion complexes have been subject to intense research during the last several decades [25, 205-208]. Due to their exceptional generic properties, macrocycles can be considered as an important class of drug delivery vehicles, able to accommodate drugs within their structure thus affording a steric barrier to drug degradation and/or deactivation. Moreover, the size of a macrocycle can be tailored to control the rate of drug release and binding strength of the host-guest complex [207, 209-212]. A number of important types of macrocyclic molecular hosts have been developed and characterized, including crown ethers [213], cyclodextrins [207], calix[n]arenes [208] and

cucurbit[n]urils [25]. These macrocyclic compounds offer an interior concave surface available to accommodate guest molecules such as drugs and diagnostic agents. Moreover, these macrocyclic molecular hosts can be chemically modified to allow tailoring of their physicochemical and complexation properties according to desired application [205, 212].

Cyclodextrins

Cyclodextrins (CDs) comprise a class of crystalline, nonhygroscopic, cyclic oligosaccharides derived from starch, containing at least six D-(+)-glucopyranose units attached by $(\alpha 1,4)$ glucoside bonds [207, 214]. Among the most commonly utilized members of this class are the naturally occurring a-, b-, and gcyclodextrin, which comprise respectively 6, 7, and 8 glucose units [215]. Cyclodextrins are 'bucketlike' or 'conelike' toroid molecules, with a rigid structure and a central cavity, the size of which varies according to the CD type. Based on the molecular architecture and the specific arrangement of polar hydroxyl functions the internal surface of the CD's cavity is hydrophobic, whereas the outside of the torus is hydrophilic [207]. These structural peculiarities allow the CD to accommodate a guest molecule within the cavity, forming an inclusion complex. CD interactions with drug molecules result primarily in enhancement to dissolution characteristics and bioavailability owing to enhanced solubility and improved chemical and physical stability [215]. Although it is the least soluble analog b-cyclodextrin is the most widely used agent from this class of excipients, because it is the least expensive, is commercially available from a variety of sources, and is capable to accomodate a number of guest-molecules of pharmaceutical interest.

In a new report a water-soluble trans-platinum complex was synthesized by inclusion complexation with beta-cyclodextrin. The complexation was confirmed by ¹H NMR, FT-IR, TGA, and XRD as well as by SEM and EDX. It was shown that the encapsulation with cyclodextrin allowed to solubilize the otherwise porly soluble metal compound to a solubility value of 1.6 mg/mL. Moreover, the cytotoxicity in vitro of the novel inclusion complex indicated a much higher activity after encapsulation [216].

Cucurbit[n]urils

Cucurbit[n]urils (CB[n]), comprise a relatively new family of macrocyclic molecular hosts that has exhibited promising results in improving anticancer drug delivery [24-26, 217, 218]. Partial or complete encapsulation of drugs within the homologues CB[6],

CB[7] or CB[8] can hinder untoward interactions with small molecule solutes or proteins and impart enhanced chemical and biopharmaceutical stability, with concomitant improve drug solubility and control drug release [24, 25, 217, 219]. These favorable features are concomitant with low intrinsic toxicity potential of the CB[n] family of nano-containers [24, 26]. Moreover, by means of modification with homing moieties CB[n] could be transformed from simple molecular hosts to targeted delivery systems, as recently documented for CB[6]-hyaluronate conjugates and lectin-seeking sugar-decorated CB[6] derivatives [220, 221].

CB[7] has been shown to effectively accommodate oxaliplatin with consequent enhancement of the stability of the drug. Most notably the encapsulated oxaliplatin is far less reactive to L-methionine [222] and thus the inclusion of platinum drugs in cucurbit[7]uril could be considered a vital strategy towards hindering untoward reactions with nucleophiles, as eventually justified with other platinum metallodrugs [217].

cisplatin has been also s effectively loaded in CB[7] occupying the central cavity of the molecular host whereby platinum atom and both Cl-ligands are located inside the macrocycle [5, 223]. In vitro studies have shown that CB[7] has no effect on the in vitro cytotoxicity of cisplatin in the human ovarian carcinoma cell line A2780 and its cisplatin-resistant sub-lines A2780/cp70 and MCP1. Nevertheless the CB[7] inclusion greatly modulates the in vivo potency of the drug against human tumor xenografts. Thus while cisplatin-CB[7] is just as effective on the chemosensitive A2780 xenografts compared with free cisplatin, whilst in the cisplatin -resistant A2780/ cp70 model the inclusion complex markedly inhibited tumor growth. Delianation of the mechanistic aspects of this ability of CB to overcome resistance in vivo pointed out that this is due to a pharmacokinetic effect. Whilst the peak plasma level and tissue distribution are the same for cisplatin -CB[7] and free cisplatin, the total concentration of circulating cisplatin-CB[7] over a period of 24 hours is significantly higher than for free platinum drug when administered at the equivalent dose, suggesting for intensified exposure of malignant lesions [223]. CB[n] proved to effectively encapsulate novel preclinical multinuclear platinum complexes without greatly compromising their cytotoxicity [218, 219, 224, 225].

Calix[n]arenes

Among the macrocyclic hosts calixarenes have

received special appraisal because of their unique three dimensional structure, facile large scale synthesis and the exceptional possibilities to undergo further synthetic elaboration and functionalization [22, 208, 211, 212]. Calix[n]arenes are macrocyclic compounds composed of phenolic units linked by methylene or sulfur groups at the 2,6-positions. They encompass a "cup"-like rigid conformation, with defined lower and upper rims and a central annulus, with a central cavity large enough to accommodate small molecules and ions [208, 212]. Moreover, their chemical transformability and the possibilities for upper and lower rim modification can allow supramolecular interactions and complex-formation with larger molecules, including proteins and nucleic acids, further broadening the biomedical applicability of calixarenes [209, 210, 226]. As far as drug delivery is concerned however, despite of the numerous favorable characteristics of calix[n]arenes these share the distinction of being water insoluble, which greatly limits their practical utility. On these grounds considerable efforts have been focused on approaches to increasing the water solubility of these compounds via introduction of polar functional groups or moieties such as sulfonates [210, 227], phosphonates [228], amines and amino acids [22, 226, 229], guanidinium [230], peptides [211], saccharides [231, 232], or polyethylene oxide (PEO) [233-235], either directly or via linkers to the upper or lower rims of calix[n]arenes [212].

The encapsulation of three platinum(II)-based anticancer complexes with DNA-intercalating phenanthroline motifs in p-sulfonatocalix[4]arene (s-CX[4]) has been examined. All three metal complexes formed 2:2 inclusion complexes with s-CX[4] where the two metal complexes stacked in a head-to-tail configuration and were capped by the s-CX[4] molecules. Encapsulation of the metal complexes in either CX[4] significantly decreased the metal complexes' rate of diffusion and protected the guest molecule from degradation by reduced L-glutathione. In vitro growth inhibition assays using the LoVo human colorectal cancer cell line showed no significant change in the cytotoxicity of one of the encapsulated complexes when encapsulated by CX[4] host and hence it could be regarded as a suitable drug delivery systems for platinum coordination compounds [236].

Moreover, substituted calixarenes have been recently described as feasible drug delivery platforms for efficient accommodation of a series of cytotoxic dinuclear platinum complexes [237].

Conclusions and future prospectus

The landmark progress in our understanding of human genomics, cellular and molecular biology and their relevance to neoplastic disease during the last decades have led to unprecedented delineation of the signal-transduction pathways and their precise role in malignant transformation and tumor biology with concomitant identification of new therapeutic targets. These advances have fuelled much research efforts upon the rational design of targeted, patient-friendly anticancer drugs. While the undisputable advantages these innovative strategies pose, relative to the classical chemotherapeutic armamentarium, they are by no means devoid of limitations, concerning their limited capacity to eradicate malignant populations, the emergence of resistance with monotherapy and their potential for pharmacokinetic drug-drug interactions.

Thus to this end the conventional "heavy-duty" anticancer drugs appear to be an inevitable component of cancer management. On these grounds the opportunities offered by state-of-the-art delivery technologies in terms of profound improvement of the biodistribution and toxicity profiles of existing drugs condition the exceptional and long-standing research efforts towards formulation design for targeted delivery of chemotherapeutics, including cisplatin and its congeners.

In line with the unique characteristics of the nanosized drug delivery systems significant development has been reported on development of nanopharmaceutical platforms for cisplatin and other platinum complexes. Using surface modification with polymers and/ or homing fragments or chemical functionalization it is feasible to tailor the generic properties of nano-scale objects such as micelles, liposomes, macromolecular pro-drugs, supramolecular complexes etc. for optimal targeting and remote drug release. Nevertheless, the elaboration of these sophisticated delivery strategies is associated with different problems such as hampered drug release, and unfortunate cost-effectiveness, which have hampered the progress of the field beyond the experimental stages, and recall for further more detailed characterization of nanocarriers as possible delivery vehicles for platinum drugs.

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References

- 1. Desoize B, Madoulet C. Particular aspects of platinum compounds used at present in cancer treatment. Crit Rev Oncol Hematol 2002;42(3):317-25.
- 2. Wheate NJ, Walker S, Craig GE, Oun R. The status of platinum anticancer drugs in the clinic and in clinical trials. Dalton Trans 2010;39(35):8113-27.
- 3. Chabner BA, Amrein PC, Druker BJ, Michaelson D, Mitsiades CS, Goss PE, et al. Chemotherapy of neoplastic disease. In: Brunton LL, Lazo JS, Parker KL, eds. Goodman & Gilman's The pharmacological basis of therapeutics. 11 ed. New York: McGraw Hill 2006:1315 403.
- 4. Momekov G, Bakalova A, Karaivanova M. Novel approaches towards development of non-classical platinum-based antineoplastic agents: Design of platinum complexes characterized by an alternative DNA-binding pattern and/or tumor-targeted cytotoxicity. Curr Med Chem 2005;12(19):2177-91.
- 5. Momekov G, Momekova D. Recent developments in antitumor platinum coordination compounds. Expert Opinion on Therapeutic Patents 2006;16(10):1383-403.
- 6. Reich SD. Toxicity of anticancer drugs used in children. Cancer Nurs 1980;3(5):385-6.
- 7. Powis G. Dose-dependent metabolism, therapeutic effect, and toxicity of anticancer drugs in man. Drug Metab Rev 1983;14(6):1145-63.
- 8. Kelland L. The resurgence of platinum-based cancer chemotherapy. Nat Rev Cancer 2007;7(8):573-84.
- 9. Ota K. [Nedaplatin]. Gan To Kagaku Ryoho 1996;23(3):379-87.
- 10. M c K e a g e MJ. Lobaplatin: a new antitumour platinum drug. Expert Opin Investig Drugs 2001;10(1):119-28.
- 11. Ahn JH, Kang YK, Kim TW, Bahng H, Chang HM, Kang WC, et al. Nephrotoxicity of heptaplatin: a randomized comparison with cisplatin in advanced gastric cancer. Cancer Chemother Pharmacol 2002;50(2):104-10.
- Min YJ, Bang SJ, Shin JW, Kim DH, Park JH, Kim GY, et al. Combination chemotherapy with 5-fluorouracil and heptaplatin as first-line treatment in patients with advanced gastric cancer. J Korean Med Sci 2004;19(3):369-73.

- 13. Ahn MJ, Oh HS, Choi JH, Lee YY, Kim IS, Choi IY, et al. Combination chemotherapy of heptaplatin, paclitaxel and 5-fluorouracil in patients with advanced gastric cancer: a pilot study. Cancer Res Treat 2004;36(3):182-6.
- 14. Lee JW, Park JK, Lee SH, Kim SY, Cho YB, Kuh HJ. Anti-tumor activity of heptaplatin in combination with 5-fluorouracil or paclitaxel against human head and neck cancer cells in vitro. Anticancer Drugs 2006;17(4):377-84.
- 15. Storm G, Crommelin DJA. Liposomes: quo vadis? PSST 1998;1(1):19-31.
- 16. Drummond DC, Meyer O, Hong K, Kirpotin DB, Papahadjopoulos D. Optimizing liposomes for delivery of chemotherapeutic agents to solid tumors. Pharmacol Rev 1995;51(4):691-743.
- 17. Drummond DC, Zignani M, Leroux J-C. Current status of pH-sensitive liposomes in drug delivery. Prog Lipid Res 2000;29:409-60.
- 18. Bildstein L, Dubernet C, Couvreur P. Prodrug-based intracellular delivery of anticancer agents. Adv Drug Deliv Rev 2011;63(1-2):3-23.
- 19. Kratz F, Muller IA, Ryppa C, Warnecke A. Prodrug strategies in anticancer chemotherapy. ChemMedChem 2008;3(1):20-53.
- 20. Deng C, Jiang Y, Cheng R, Meng F, Zhong Z. Biodegradable polymeric micelles for targeted and controlled anticancer drug delivery: Promises, progress and prospects. Nano Today 2012; In press.
- 21. M a D, Hettiarachchi G, Nguyen D, Zhang B, Wittenberg JB, Zavalij PY, et al. Acyclic cucurbit[n]uril molecular containers enhance the solubility and bioactivity of poorly soluble pharmaceuticals. Nat Chem 2012;4(6):503-10.
- 22. Bew SP, Brimage RA, L'Hermite N, Sharma SV. Upper rim appended hybrid calixarenes via click chemistry. Org Lett 2007;9(19):3713-6.
- 23. Da Silva E, Lazar AN, Coleman AW. Biopharmaceutical applications of calixarenes. J Drug Deliv Sci Tech 2004;14:3-20.
- 24. Hettiarachchi G, Nguyen D, Wu J, Lucas D, Ma D, Isaacs L, et al. Toxicology and drug delivery by cucurbit[n]uril type molecular containers. PLoS One 2010:1-10.
- 25. Lagona J, Mukhopadhyay P, Chakrabarti S, Isaacs L. The cucurbit[n]uril family. Angew Chem Int Ed Engl 2005;44(31):4844-70.

- 26. Walker S, Oun R, McInnes FJ, Wheate NJ. The potential of cucurbit[n]urils in drug delivery. Isr J Chem 2011;51(5-6):616-24.
- 27. Riggio C, Pagni E, Raffa V, Cuschieri A. Nano-oncology: clinical application for cancer therapy and future perspectives. J Nanomaterials 2011:1-10.
- 28. Brewer E, Coleman J, Lowman A. Emerging technologies of polymeric nanoparticles in cancer drug delivery. J Nanomaterials 2011:1-10.
- 29. A1-Jamal WT, Kostarelos K. Liposomes: from a clinically established drug delivery system to a nanoparticle platform for theranostic nanomedicine. Acc Chem Res 2011;44(10):1094-104.
- 30. Jolck RI, Feldborg LN, Andersen S, Moghimi SM, Andresen TL. Engineering liposomes and nanoparticles for biological targeting. Adv Biochem Eng Biotechnol 2011;125:251-80.
- 31. Langer M, Beck-Sickinger AG. Peptides as carrier for tumor diagnosis and treatment. Curr Med Chem Anticancer Agents 2001;1(1):71-93.
- 32. Maeda H, Wu J, Sawa T, Matsumura Y, Hori K. Tumor vascular permeability and the EPR effect in macromolecular therapeutics: a review. J Control Release 2000;65(1-2):271-84.
- 33. A b d e l r a h i m M, Konduri S, Basha R, Philip PA, Baker CH. Angiogenesis: an update and potential drug approaches (review). Int J Oncol 2010;36(1):5-18.
- 34. Matsumura Y, Maeda H. A new concept for macromolecular therapeutics in cancer chemotherapy: mechanism of tumoritropic accumulation of proteins and the antitumor agent smancs. Cancer Res 1986;46(12 Pt 1):6387-92.
- 35. Konno T, Ohtsuka N, Yamasaki K, Mizutani J, Miyauchi Y, Maeda H, et al. Targeting of anticancer chemotherapy utilizing the characteristic nature of the neovasculature of solid tumors. Gan To Kagaku Ryoho 1986;13(4 Pt 2):1448-55.
- 36. Matsumura Y, Oda T, Maeda H. General mechanism of intratumor accumulation of macromolecules: advantage of macromolecular therapeutics. Gan To Kagaku Ryoho 1987;14(3 Pt 2):821-9.
- 37. Maeda H, Matsumura Y. Tumoritropic and lymphotropic principles of macromolecular drugs. Crit Rev Ther Drug Carrier Syst 1989;6(3):193-210.

- 38. Maeda H, Matsumura Y. EPR effect based drug design and clinical outlook for enhanced cancer chemotherapy. Adv Drug Deliv Rev 2011;63(3):129-30.
- 39. Moghimi SM, Szebeni J. Stealth liposomes and long circulating nanoparticles: critical issues in pharmacokinetics, opsonization and protein-binding properties. Prog Lipid Res 2003;42(6):463-78.
- 40. Metselaar JM, Mastrobattista E, Storm G. Liposomes for intravenous drug targeting: design and applications Mini Rev Med Chem 2002;2(4):319-29.
- 41. Freise J, Muller WH, Magerstedt P. Uptake of liposomes and sheep red blood cells by the liver and spleen of rats with normal or decreased function of the reticuloendothelial system. Res Exp Med (Berl) 1981;178(3):263-9.
- 42. Qi XR, Maitani Y, Nagai T. Rates of systemic degradation and reticuloendothelial system uptake of calcein in the dipalmitoylphosphatidylcholine liposomes with soybean-derived sterols in mice. Pharm Res 1995;12(1):49-52.
- 43. Castelli DD, Terreno E, Cabella C, Chaabane L, Lanzardo S, Tei L, et al. Evidence for in vivo macrophage mediated tumor uptake of paramagnetic/fluorescent liposomes. NMR Biomed 2009;22(10):1084-92.
- 44. Torchilin VP. Recent advances with liposomes as pharmaceutical carriers. Nat Rev Drug Discov 2005;4(2):145-60.
- 45. Sawant RR, Torchilin VP. Polymeric micelles: polyethylene glycol-phosphatidylethanolamine (PEG-PE)-based micelles as an example. Methods Mol Biol 2010;624:131-49.
- 46. Sawant RR, Torchilin VP. Multifunctionality of lipid-core micelles for drug delivery and tumour targeting. Mol Membr Biol 2010;27(7):232-46.
- 47. Sarisozen C, Vural I, Levchenko T, Hincal AA, Torchilin VP. PEG-PE-based micelles co-loaded with paclitaxel and cyclosporine A or loaded with paclitaxel and targeted by anticancer antibody overcome drug resistance in cancer cells. Drug Deliv 2012;19(4):169-76.
- 48. Woodle MC. Controlling liposome blood clearance by surface-grafted polymers. Adv Drug Deliv Rev 1998;32(1-2):139-52.
- 49. Chirino AJ, Ary ML, Marshall SA. Minimizing the immunogenicity of protein therapeutics. Drug Discov Today 2004;9(2):82-90.

- 50. Elizondo E, Moreno E, Cabrera I, Cordoba A, Sala S, Veciana J, et al. Liposomes and other vesicular systems: structural characteristics, methods of preparation, and use in nanomedicine. Prog Mol Biol Transl Sci 2011;104:1-52.
- 51. Hilger I, Hergt R, Kaiser WA. Use of magnetic nanoparticle heating in the treatment of breast cancer. IEE Proc Nanobiotechnol 2005;152(1):33-9.
- 52. Sampathkumar SG, Yarema KJ. Targeting cancer cells with dendrimers. Chem Biol 2005;12(1):5-6.
- 53. Thomas TP, Choi SK, Li MH, Kotlyar A, Baker JR, Jr. Design of riboflavin-presenting PAMAM dendrimers as a new nanoplatform for cancer-targeted delivery. Bioorg Med Chem Lett 2010;20(17):5191-4.
- 54. Yellepeddi VK, Kumar A, Palakurthi S. Biotinylated poly(amido)amine (PAMAM) dendrimers as carriers for drug delivery to ovarian cancer cells in vitro. Anticancer Res 2009;29(8):2933-43.
- 55. Yellepeddi VK, Kumar A, Maher DM, Chauhan SC, Vangara KK, Palakurthi S. Biotinylated PAMAM dendrimers for intracellular delivery of cisplatin to ovarian cancer: role of SMVT. Anticancer Res 2011;31(3):897-906.
- 56. Soininen SK, Lehtolainen-Dalkilic P, Karppinen T, Puustinen T, Dragneva G, Kaikkonen MU, et al. Targeted delivery via avidin fusion protein: Intracellular fate of biotinylated doxorubicin derivative and cellular uptake kinetics and biodistribution of biotinylated liposomes. Eur J Pharm Sci 2012.
- 57. Sun M, Wang Y, Shen J, Xiao Y, Su Z, Ping Q. Octreotide-modification enhances the delivery and targeting of doxorubicin-loaded liposomes to somatostatin receptors expressing tumor in vitro and in vivo. Nanotechnology 2010:475101.
- 58. Zhang Y, Zhang H, Wang X, Wang J, Zhang X, Zhang Q. The eradication of breast cancer and cancer stem cells using octreotide modified paclitaxel active targeting micelles and salinomycin passive targeting micelles. Biomaterials 2011;33(2):679-91.
- 59. Paliwal SR, Paliwal R, Pal HC, Saxena AK, Sharma PR, Gupta PN, et al. Estrogenanchored pH-sensitive liposomes as nanomodule designed for site-specific delivery of doxo-

- rubicin in breast cancer therapy. Mol Pharm 2012;9(1):176-86.
- 60. He Y, Zhang L, Song C. Luteinizing hormonereleasing hormone receptor-mediated delivery of mitoxantrone using LHRH analogs modified with PEGylated liposomes. Int J Nanomedicine 2010;5:697-705.
- 61. Singh M. Transferrin as a targeting ligand for liposomes and anticancer drugs. Curr Pharm Des 1999;5(6):443-51.
- 62. Dass CR, Choong PF. Targeting of small molecule anticancer drugs to the tumour and its vasculature using cationic liposomes: lessons from gene therapy. Cancer Cell Int 2006;6:17.
- 63. Wang S, XuH, XuJ, Zhang Y, LiuY, Deng YH, et al. Sustained liver targeting and improved antiproliferative effect of doxorubicin liposomes modified with galactosylated lipid and PEGlipid. AAPS PharmSciTech 2010;11(2):870-7.
- 64. Yang R, Meng F, MaS, Huang F, Liu H, Zhong Z. Galactose-decorated cross-linked biodegradable poly(ethylene glycol)-b-poly(epsilon-caprolactone) block copolymer micelles for enhanced hepatoma-targeting delivery of paclitaxel. Biomacromolecules 2011;12(8):3047-55.
- 65. Zhao H, Wang JC, Sun QS, Luo CL, Zhang Q. RGD-based strategies for improving antitumor activity of paclitaxel-loaded liposomes in nude mice xenografted with human ovarian cancer. J Drug Target 2009;17(1):10-8.
- 66. Jiang J, Yang SJ, Wang JC, Yang LJ, Xu ZZ, Yang T, et al. Sequential treatment of drug-resistant tumors with RGD-modified liposomes containing siRNA or doxorubicin. Eur J Pharm Biopharm 2010;76(2):170-8.
- 67. Dai W, Yang T, Wang X, Wang J, Zhang X, Zhang Q. PHSCNK-Modified and doxorubicin-loaded liposomes as a dual targeting system to integrin-overexpressing tumor neovasculature and tumor cells. J Drug Target 2010;18(4):254-63.
- 68. Meng S, Su B, Li W, Ding Y, Tang L, Zhou W, et al. Enhanced antitumor effect of novel dual-targeted paclitaxel liposomes. Nanotechnology 2010:415103.
- 69. Cao Z, Tong R, Mishra A, Xu W, Wong GC, Cheng J, et al. Reversible cell-specific drug delivery with aptamer-functionalized liposomes. Angew Chem Int Ed Engl 2009;48(35):6494-8.
- 70. Huang YF, Shangguan D, Liu H, Phillips JA, Zhang X, Chen Y, et al. Molecular assem-

- bly of an aptamer-drug conjugate for targeted drug delivery to tumor cells. Chembiochem 2009;10(5):862-8.
- 71. Zhang Y, Hong H, Cai W. Tumor-targeted drug delivery with aptamers. Curr Med Chem 2011;18(27):4185-94.
- 72. Dhar S, Gu FX, Langer R, Farokhzad OC, Lippard SJ. Targeted delivery of cisplatin to prostate cancer cells by aptamer functionalized Pt(IV) prodrug-PLGA-PEG nanoparticles. Proc Natl Acad Sci U S A 2008;105(45):17356-61.
- 73. Dhar S, Kolishetti N, Lippard SJ, Farokhzad OC. Targeted delivery of a cisplatin prodrug for safer and more effective prostate cancer therapy in vivo. Proc Natl Acad Sci U S A 2011;108(5):1850-5.
- 74. Slepushkin V, Simoes S, de Lima MC, Duzgunes N. Sterically stabilized phsensitive liposomes. Methods Enzymol 2004;387:134-47.
- 75. Salmaso S, Bersani S, Pirazzini M, Caliceti P. pH-sensitive PEG-based micelles for tumor targeting. J Drug Target 2010;19(4):303-13.
- 76. Chen W, Meng F, Cheng R, Zhong Z. pH-Sensitive degradable polymersomes for triggered release of anticancer drugs: a comparative study with micelles. J Control Release 2010;142(1):40-6.
- 77. Vanic Z, Barnert S, Suss R, Schubert R. Fusogenic activity of PEGylated pH-sensitive liposomes. J Liposome Res 2011;22(2):148-57.
- 78. Tannock IF, Rotin D. Acid pH in tumors and its potential for therapeutic exploitation. Cancer Res 1989;49(16):4373-84.
- 79. Slepushkin VA, Simoes S, Dazin P, Newman MS, Guo LS, Pedroso de Lima MC, et al. Sterically stabilized pH-sensitive liposomes. Intracellular delivery of aqueous contents and prolonged circulation in vivo. J Biol Chem 1997;272(4):2382-8.
- 80. Simoes S, Slepushkin V, Duzgunes N, Pedroso de Lima MC. On the mechanisms of internalization and intracellular delivery mediated by pH-sensitive liposomes. Biochim Biophys Acta 2001;1515(1):23-37.
- 81. Morilla MJ, Romero EL. Liposomal pH-sensitive nanomedicines in preclinical development. In: Reisner DE, ed. Bionanotechnology II: Global Prospects. Boca Raton: CRC-Press 2011:383-413.

- 82. Bibi S, Lattmann E, Mohammed AR, Perrie Y. Trigger release liposome systems: local and remote controlled delivery? J Microencapsul 2012;29(3):262-76.
- 83. Bergstrand N, Arfvidsson MC, Kim JM, Thompson DH, Edwards K. Interactions between pH-sensitive liposomes and model membranes. Biophys Chem 2003;104(1):361-79.
- 84. New RRC. Introduction. In: New RRC, ed. Liposomes: a practical approach. Oxford: Oxford University Press 1994:1-32.
- 85. Andresen TL, Jensen SS, Jorgensen K. Advanced strategies in liposomal cancer therapy: problems and prospects of active and tumor specific drug release. Prog Lipid Res 2005;44(1):68-97.
- 86. Lasic DD, Vallner JJ, Working PK. Sterically stabilized liposomes in cancer therapy and gene delivery. Curr Opin Mol Ther 1999;1(2):177-85.
- 87. Andresen TL, Jensen SS, Kaasgaard T, Jorgensen K. Triggered activation and release of liposomal prodrugs and drugs in cancer tissue by secretory phospholipase A2. Curr Drug Deliv 2005;2(4):353-62.
- 88. Lasic D, Martin F, eds. Stealth Liposomes. Boca Raton: CRC Press 1995.
- 89. Kelland L. Broadening the clinical use of platinum drug-based chemotherapy with new analogues. Satraplatin and picoplatin. Expert Opin Investig Drugs 2007;16(7):1009-21.
- 90. Kaluderovic GN, Dietrich A, Kommera H, Kuntsche J, Mader K, Mueller T, et al. Liposomes as vehicles for water insoluble platinum-based potential drug: 2-(4-(Tetrahydro-2H-pyran-2-yloxy)-undecyl)-propane-1,3-diamminedichloroplatinum(II). Eur J Med Chem 2012;54:567-72.
- 91. Storm G. Liposomes as delivery system for doxorubicin in cancer chemotherapy. Pharm Weekbl Sci 1988;10(6):288-90.
- 92. Storm G, Wilms HP, Crommelin DJ. Liposomes and biotherapeutics. Biotherapy 1991;3(1):25-42.
- 93. Zeisig R, Shimada K, Hirota S, Arndt D. Effect of sterical stabilization on macrophage uptake in vitro and on thickness of the fixed aqueous layer of liposomes made from alkylphosphocholines. Biochim Biophys Acta 1996;1285(2):237-45.

- 94. Fedier A, Poyet C, Perucchini D, Boulikas T, Fink D. MLH1-deficient tumor cells are resistant to lipoplatin, but retain sensitivity to lipoxal. Anticancer Drugs 2006;17(3):315-23.
- 95. Charest G, Paquette B, Fortin D, Mathieu D, Sanche L. Concomitant treatment of F98 glioma cells with new liposomal platinum compounds and ionizing radiation. J Neurooncol 2010;97(2):187-93.
- 96. Charest G, Sanche L, Fortin D, Mathieu D, Paquette B. Glioblastoma treatment: bypassing the toxicity of platinum compounds by using liposomal formulation and increasing treatment efficiency with concomitant radiotherapy. Int J Radiat Oncol Biol Phys 2012;84(1):244-9.
- 97. Carvalho Junior AD, Vieira FP, Melo VJ, Lopes MT, Silveira JN, Ramaldes GA, et al. Preparation and cytotoxicity of cisplatin-containing liposomes. Braz J Med Biol Res 2007;40(8):1149-57.
- 98. Peleg-Shulman T, Gibson D, Cohen R, Abra R, Barenholz Y. Characterization of sterically stabilized cisplatin liposomes by nuclear magnetic resonance. Biochim Biophys Acta 2001;1510(1-2):278-91.
- 99. Va a g e J, Donovan D, Wipff E, Abra R, Colbern G, Uster P, et al. Therapy of a xenografted human colonic carcinoma using cisplatin or doxorubicin encapsulated in long-circulating pegylated stealth liposomes. Int J Cancer 1999;80(1):134-7.
- 100. Chang H-I, Yeh M-K. Clinical development of liposome-based drugs: formulation, characterization, and therapeutic efficacy. Int J Nanomedicine 2012;7:49–60.
- 101. Qian S, Li C, Zuo Z. Pharmacokinetics and disposition of various drug loaded liposomes. Curr Drug Metab 2012;13(4):372-95.
- 102. B o u l i k a s T. Clinical overview on Lipoplatin: a successful liposomal formulation of cisplatin. Expert Opin Investig Drugs 2009;18(8):1197-218.
- 103. Jehn CF, Boulikas T, Kourvetaris A, Kofla G, Possinger K, Luftner D. First safety and response results of a randomized phase III study with liposomal platin in the treatment of advanced squamous cell carcinoma of the head and neck (SCCHN). Anticancer Res 2008;28(6B):3961-4.
- 104. Stathopoulos GP. Liposomal cisplatin: a new cisplatin formulation. Anticancer Drugs 2010;21(8):732-6.

- 105. Stathopoulos GP, Boulikas T, Kourvetaris A, Stathopoulos J. Liposomal oxaliplatin in the treatment of advanced cancer: a phase I study. Anticancer Res 2006;26(2B):1489-93.
- 106. A rienti C, Tesei A, Ravaioli A, Ratta M, Carloni S, Mangianti S, et al. Activity of lipoplatin in tumor and in normal cells in vitro. Anticancer Drugs 2008;19(10):983-90.
- 107. R a v a i o l i A, Papi M, Pasquini E, Marangolo M, Rudnas B, Fantini M, et al. Lipoplatin monotherapy: A phase II trial of second-line treatment of metastatic non-small-cell lung cancer. J Chemother 2009;21(1):86-90.
- 108. Canta A, Chiorazzi A, Carozzi V, Meregalli C, Oggioni N, Sala B, et al. In vivo comparative study of the cytotoxicity of a liposomal formulation of cisplatin (lipoplatin). Cancer Chemother Pharmacol 2011;68(4):1001-8.
- 109. Stathopoulos GP, Antoniou D, Dimitroulis J, Stathopoulos J, Marosis K, Michalopoulou P. Comparison of liposomal cisplatin versus cisplatin in non-squamous cell non-small-cell lung cancer. Cancer Chemother Pharmacol 2011;68(4):945-50.
- 110. Storm G, Steerenberg PA, Emmen F, van Borssum Waalkes M, Crommelin DJ. Release of doxorubicin from peritoneal macrophages exposed in vivo to doxorubicin-containing liposomes. Biochim Biophys Acta 1988;965(2-3):136-45.
- 111. Storm G, Van Gessel HJ, Steerenberg PA, Speth PA, Roerdink FH, Regts J, et al. Investigation of the role of mononuclear phagocytes in the transportation of doxorubicin-containing liposomes into a solid tumor. Cancer Drug Deliv 1987;4(2):89-104.
- 112. Storm G, ten Kate MT, Working PK, Bakker-Woudenberg IA. Doxorubicin entrapped in sterically stabilized liposomes: effects on bacterial blood clearance capacity of the mononuclear phagocyte system. Clin Cancer Res 1998;4(1):111-5.
- 113. S z e b e n i J, Baranyi L, Savay S, Milosevits J, Bunger R, Laverman P, et al. Role of complement activation in hypersensitivity reactions to doxil and hynic PEG liposomes: experimental and clinical studies. J Liposome Res 2002;12(1-2):165-72.
- 114. Li C, Wang C, Yang H, Zhao X, Wei N, Cui J. Liposomal topotecan formulation with a low

- polyethylene glycol grafting density: pharmacokinetics and antitumour activity. J Pharm Pharmacol 2012;64(3):372-82.
- 115. Li CL, Cui JX, Wang CX, Zhang L, Li YH, Zhang L, et al. Development of pegylated liposomal vinorelbine formulation using "post-insertion" technology. Int J Pharm 2010;391(1-2):230-6.
- 116. Li C, Cui J, Wang C, Zhang L, Xiu X, Li Y, et al. Encapsulation of vinorelbine into cholesterol-polyethylene glycol coated vesicles: drug loading and pharmacokinetic studies. J Pharm Pharmacol 2011;63(3):376-84.
- 117. Momekov a D, Momekov G, Rangelov S, Storm G, Lambov N. Physicochemical and bi-opharmaceutical characterization of dipalmitoyl phosphatidylcholine liposomes sterically stabilized by copolymers bearing short blocks of lipid-mimetic units. Soft Matter 2010;6(3):59-601.
- 118. Momekova D, Rangelov S, Lambov N. Preparation and properties of soybean phosphatidylcholine liposomes sterically stabilized by copolymers bearing short blocks of lipid-mimetic units Compt Rend Acad Bulg Sci 2007;60(7):769-74.
- 119. Momekova D, Rangelov S, Lambov N, Karlsson G, Almgren M. Effects of amphiphilic copolymers bearing short blocks of lipid-mimetic units on the membrane properties and morphology of DSPC liposomes. J Disp Sci Technol 2008;29:1106-13.
- 120. Romberg B, Hennink WE, Storm G. Sheddable coatings for long-circulating nanoparticles. Pharm Res 2008;25(1):55-71.
- 121. Romberg B, Metselaar JM, Baranyi L, Snel CJ, Bunger R, Hennink WE, et al. Poly(amino acid)s: promising enzymatically degradable stealth coatings for liposomes. Int J Pharm 2007;331(2):186-9.
- 122. Romberg B, Metselaar JM, deVringer T, Motonaga K, Kettenes-van den Bosch JJ, Oussoren C, et al. Enzymatic degradation of liposome-grafted poly(hydroxyethyl L-glutamine). Bioconjug Chem 2005;16(4):767-74.
- 123. Romberg B, Oussoren C, Snel CJ, Carstens MG, Hennink WE, Storm G. Pharmacokinetics of poly(hydroxyethyl-l-asparagine)-coated liposomes is superior over that of PEG-coated liposomes at low lipid dose and upon repeated administration. Biochim Biophys Acta 2007;1768(3):737-43.

- 124. Kenworthy AK, Hristova K, Neetham D, McIntosh TJ. Range and magnitude of the steric pressure between bilayers containing phospholipids with covalently attached poly(ethylene glycol). Biophys J 1995;68:1921-36.
- 125. Hristova K, Kenworthy A, McIntosh TJ. Effect of bilayer composition on the phase behaviour of liposomal suspensions containing poly(ethylene glycol)-lipids. Macromolecules 1995;28:7693-9.
- 126. Marsh D, Bartucci R, Sportelli L. Lipid membranes with grafted polymers: physicochemical aspects. Biochim Biophys Acta 2003 1615:33-59.
- 127. R angelov S, Edwards K, Almgren M, Karlsson G. Steric stabilization of egg-phosphathidyl choline liposomes by copolymers bearing short blocks of lipid-mimetic units. Langmuir 2003; 19:172–81.
- 128. Rangelov S, Petrova E, Berlinova I, Tsevetanov C. Synthesis and polymerization of novel oxirane bearing an aliphatic double chain moiety. Polymer 2001;42:4483–91.
- 129. Momekova D, Rangelov S, Lambov N. Long-circulating, pH-sensitive liposomes. Methods Mol Biol 2010;605:527-44.
- 130. Momekova D, Rangelov S, Yanev S, Nikolova E, Konstantinov S, Romberg B, et al. Long-circulating, pH-sensitive liposomes sterically stabilized by copolymers bearing short blocks of lipid-mimetic units. Eur J Pharm Sci 2007;32(4-5):308-17.
- 131. Momekova D, Momekov G, Rangelov S, Lambov N. In vitro biocompatibility study of free and liposomaly-grafted copolymers bearing short blocks of aliphatic lipid-mimetic units cytotoxicity and hemolytic activity. J Drug Deliv Sci Technol 2007;17(6):393-7.
- 132. Momekova D, Momekov G, Pencheva I, Rangelov S, Lambov N. Formulation of bendamustine hydrochloride in long circulating DPPC:CHOL liposomes, surface modified with a PEO-based co-polymer bearing four lipid mimetic units. Pharmacia 2012;In press.
- 133. Li SD, Huang L. Stealth nanoparticles: high density but sheddable PEG is a key for tumor targeting. J Control Release 2010;145(3):178–81.
- 134. Bitounis D, Fanciullino R, Iliadis A, Ciccolini J. Optimizing druggability through liposomal formulations: new approaches to an old concept. ISRN Pharmaceutics 2012:1-11.

- 135. Romberg B, Flesch FM, Hennink WE, Storm G. Enzyme-induced shedding of a poly(amino acid)-coating triggers contents release from dioleoyl phosphatidylethanolamine liposomes. Int J Pharm 2008;355(1-2):108-13.
- 136. Romberg B, Oussoren C, Snel CJ, Hennink WE, Storm G. Effect of liposome characteristics and dose on the pharmacokinetics of liposomes coated with poly(amino acid)s. Pharm Res 2007;24(12):2394-401.
- 137. Zhang YF, Wang JC, Bian DY, Zhang X, Zhang Q. Targeted delivery of RGD-modified liposomes encapsulating both combretastatin A-4 and doxorubicin for tumor therapy: in vitro and in vivo studies. Eur J Pharm Biopharm 2010;74(3):467-73.
- 138. Garg A, Kokkoli E. pH-Sensitive PEGylated liposomes functionalized with a fibronectin-mimetic peptide show enhanced intracellular delivery to colon cancer cell. Curr Pharm Biotechnol 2011;12(8):1135-43.
- 139. Naik S, Patel D, Chuttani K, Mishra AK, Misra A. In vitro mechanistic study of cell death and in vivo performance evaluation of RGD grafted PEGylated docetaxel liposomes in breast cancer. Nanomedicine 2012;8(6):951-62.
- 140. Abu Lila AS, Kizuki S, Doi Y, Suzuki T, Ishida T, Kiwada H. Oxaliplatin encapsulated in PEG-coated cationic liposomes induces significant tumor growth suppression via a dual-targeting approach in a murine solid tumor model. J Control Release 2009;137(1):8-14.
- 141. Du J, Lu WL, Ying X, Liu Y, Du P, Tian W, et al. Dual-targeting topotecan liposomes modified with tamoxifen and wheat germ agglutinin significantly improve drug transport across the blood-brain barrier and survival of brain tumorbearing animals. Mol Pharm 2009;6(3):905-17.
- 142. Ying X, Wen H, Lu WL, Du J, Guo J, Tian W, et al. Dual-targeting daunorubicin liposomes improve the therapeutic efficacy of brain glioma in animals. J Control Release 2010;141(2):183-92.
- 143. Corley P, Loughrey HC. Targeting of doxorubicin loaded liposomes to T-cells via the transferrin receptor. Biochem Soc Trans 1998;26(1):S37.
- 144. Gijsens A, Derycke A, Missiaen L, De Vos D, Huwyler J, Eberle A, et al. Targeting of the photocytotoxic compound AlPcS4 to Hela cells by transferrin conjugated PEG-liposomes. Int J Cancer 2002;101(1):78-85.

- 145. Derycke AS, De Witte PA. Transferrinmediated targeting of hypericin embedded in sterically stabilized PEG-liposomes. Int J Oncol 2002;20(1):181-7.
- 146. Lopez-Barcons LA, Polo D, Llorens A, Reig F, Fabra A. Targeted adriamycin delivery to MXT-B2 metastatic mammary carcinoma cells by transferrin liposomes: effect of adriamycin ADR-to-lipid ratio. Oncol Rep 2005;14(5):1337-43.
- 147. Yamada Y, Shinohara Y, Kakudo T, Chaki S, Futaki S, Kamiya H, et al. Mitochondrial delivery of mastoparan with transferrin liposomes equipped with a pH-sensitive fusogenic peptide for selective cancer therapy. Int J Pharm 2005;303(1-2):1-7.
- 148. An abousi S, Bakowsky U, Schneider M, Huwer H, Lehr CM, Ehrhardt C. In vitro assessment of transferrin-conjugated liposomes as drug delivery systems for inhalation therapy of lung cancer. Eur J Pharm Sci 2006;29(5):367-74.
- 149. Soni V, Kohli DV, Jain SK. Transferrin coupled liposomes as drug delivery carriers for brain targeting of 5-florouracil. J Drug Target 2005;13(4):245-50.
- 150. Li X, Ding L, Xu Y, Wang Y, Ping Q. Targeted delivery of doxorubicin using stealth liposomes modified with transferrin. Int J Pharm 2009;373(1-2):116-23.
- 151. Krieger ML, Eckstein N, Schneider V, Koch M, Royer HD, Jaehde U, et al. Overcoming cisplatin resistance of ovarian cancer cells by targeted liposomes in vitro. Int J Pharm 2010;389(1-2):10-7.
- 152. Irmer G, Burger C, Muller R, Ortmann O, Peter U, Kakar SS, et al. Expression of the messenger RNAs for luteinizing hormone-releasing hormone (LHRH) and its receptor in human ovarian epithelial carcinoma. Cancer Res 1995;55(4):817-22.
- 153. Mai J, Song S, Rui M, Liu D, Ding Q, Peng J, et al. A synthetic peptide mediated active targeting of cisplatin liposomes to Tie2 expressing cells. J Control Release 2009;139(3):174-81.
- 154. El Bayoumi TA, Torchilin VP. Current trends in liposome research. Methods Mol Biol 2010;605:527-44.
- 155. Simoes S, Moreira JN, Fonseca C, Duzgunes N, de Lima MC. On the formulation of pH-sensitive liposomes with long circulation times. Adv Drug Deliv Rev 2004;56(7):947-65.

- 156. dos Santos Giuberti C, de Oliveira Reis EC, Ribeiro Rocha TG, Leite EA, Lacerda RG, Ramaldes GA, et al. Study of the pilot production process of long-circulating and pH-sensitive liposomes containing cisplatin. J Liposome Res 2011;21(1):60-9.
- 157. Leite EA, Giuberti Cdos S, Wainstein AJ, Wainstein AP, Coelho LG, Lana AM, et al. Acute toxicity of long-circulating and pH-sensitive liposomes containing cisplatin in mice after intraperitoneal administration. Life Sci 2009;84(19-20):641-9.
- 158. Leite EA, Lana AM, Junior AD, Coelho LG, De Oliveira MC. Acute toxicity study of cisplatin loaded long-circulating and pH-sensitive liposomes administered in mice. J Biomed Nanotechnol 2012;8(2):229-39.
- 159. de Carvalho Maroni L, de Oliveira Silveira AC, Leite EA, Melo MM, de Carvalho Ribeiro AF, Cassali GD, et al. Antitumor effectiveness and toxicity of cisplatin-loaded long-circulating and pH-sensitive liposomes against Ehrlich ascitic tumor. Exp Biol Med (Maywood) 2012;237(8):973-84.
- 160. Momekov G, Momekova D, Rangelov S, Lambov N. Cytotoxicity and cellular accumulation of a novel sterically stabilized DOPE:CHEMs-based pH-sensitive liposomal formulation of mitomycin C. Pharmacia 2012;In press.
- 161. Ta T, Convertine AJ, Reyes CR, Stayton PS, Porter TM. Thermosensitive liposomes modified with poly(N-isopropylacrylamide-copropylacrylic acid) copolymers for triggered release of doxorubicin. Biomacromolecules 2010;11(8):1915-20.
- 162. Banerjee S, Sen K, Pal TK, Guha SK. Poly(styrene-co-maleic acid)-based pH-sensitive liposomes mediate cytosolic delivery of drugs for enhanced cancer chemotherapy. Int J Pharm 2012;436(1-2):786-97.
- 163. Soares DC, Cardoso VN, de Barros AL, de Souza CM, Cassali GD, de Oliveira MC, et al. Antitumoral activity and toxicity of PEG-coated and PEG-folate-coated pH-sensitive liposomes containing 159Gd-DTPA-BMA in Ehrlich tumor bearing mice. Eur J Pharm Sci 2012;45(1-2):58-64.
- 164. Andresen TL, Davidsen J, Begtrup M, Mouritsen OG, Jorgensen K. Enzymatic release

- of antitumor ether lipids by specific phospholipase A2 activation of liposome-forming prodrugs. J Med Chem 2004;47(7):1694-703.
- 165. Andresen TL, Jensen SS, Madsen R, Jorgensen K. Synthesis and biological activity of anticancer ether lipids that are specifically released by phospholipase A2 in tumor tissue. J Med Chem 2005;48(23):7305-14.
- 166. And resen TL, Jorgensen K. Synthesis and membrane behavior of a new class of unnatural phospholipid analogs useful as phospholipase A2 degradable liposomal drug carriers. Biochim Biophys Acta 2005;1669(1):1-7.
- 167. Davidsen J, Jorgensen K, Andresen TL, Mouritsen OG. Secreted phospholipase A2 as a new enzymatic trigger mechanism for localised liposomal drug release and absorption in diseased tissue. Biochim Biophys Acta 2003;1609(1):95-101.
- 168. Jensen SS, Andresen TL, Davidsen J, Hoyrup P, Shnyder SD, Bibby MC, et al. Secretory phospholipase A2 as a tumor-specific trigger for targeted delivery of a novel class of liposomal prodrug anticancer etherlipids. Mol Cancer Ther 2004;3(11):1451-8.
- 169. Linderoth L, Andresen TL, Jorgensen K, Madsen R, Peters GH. Molecular basis of phospholipase A2 activity toward phospholipids with sn-1 substitutions. Biophys J 2008;94(1):14-26.
- 170. Linderoth L, Peters GH, Jorgensen K, Madsen R, Andresen TL. Synthesis of sn-1 functionalized phospholipids as substrates for secretory phospholipase A2. Chem Phys Lipids 2007;146(1):54-66.
- 171. Peters GH, Moller MS, Jorgensen K, Ronnholm P, Mikkelsen M, Andresen TL. Secretory phospholipase A2 hydrolysis of phospholipid analogues is dependent on water accessibility to the active site. J Am Chem Soc 2007;129(17):5451-61.
- 172. liplasome.com. [homepage on the internet] [cited 2012; Available from: http://www.liplasome.com
- 173. Kaasgaard T, Andresen TL, Jensen SS, Holte RO, Jensen LT, Jorgensen K. Liposomes containing alkylated methotrexate analogues for phospholipase A(2) mediated tumor targeted drug delivery. Chem Phys Lipids 2009;157(2):94-103.

- 174. Pedersen PJ, Adolph SK, Subramanian AK, Arouri A, Andresen TL, Mouritsen OG, et al. Liposomal formulation of retinoids designed for enzyme triggered release. J Med Chem 2010;53(9):3782-92.
- 175. Evjen TJ, Hagtvet E, Nilssen EA, Brandl M, Fossheim SL. Sonosensitive dioleoylphosphatidylethanolamine-containing liposomes with prolonged blood circulation time of doxorubicin. Eur J Pharm Sci 2011;43(4):318-24.
- 176. Schroeder A, Honen R, Turjeman K, Gabizon A, Kost J, Barenholz Y. Ultrasound triggered release of cisplatin from liposomes in murine tumors. J Control Release 2009;137(1):63-8.
- 177. Li G, Liu J, Pang Y, Wang R, Mao L, Yan D, et al. Polymeric micelles with water-insoluble drug as hydrophobic moiety for drug delivery. Biomacromolecules 2011;12(6):2016-26.
- 178. Mufamadi MS, Pillay V, Choonara YE, Du Toit LC, Girish Modi G, Dinesh Naidoo D, et al. A review on composite liposomal technologies for specialized drug delivery. J Drug Deliv 2011:1-19.
- 179. Avendaño C, Menéndez JC. Medicinal Chemistry of Anticancer Drugs. Amsterdam: Elsevier 2008.
- 180. Yokoyama M, Okano T, Sakurai Y, Ekimoto H, Shibazaki C, Kataoka K. Toxicity and antitumor activity against solid tumors of micelle-forming polymeric anticancer drug and its extremely long circulation in blood. Cancer Res 1991;51(12):3229-36.
- 181. N i s h i y a m a N, Kataoka K. Preparation and characterization of size-controlled polymeric micelle containing cisdichlorodiammineplatinum(II) in the core. J Control Release 2001;74(1-3):83-94.
- 182. N i s h i y a m a N, Kato Y, Sugiyama Y, Kataoka K. Cisplatin-loaded polymer-metal complex micelle with time-modulated decaying property as a novel drug delivery system. Pharm Res 2001;18(7):1035-41.
- 183. Nishiyama N, Okazaki S, Cabral H, Miyamoto M, Kato Y, Sugiyama Y, et al. Novel cisplatin-incorporated polymeric micelles can eradicate solid tumors in mice. Cancer Res 2003;63(24):8977-83.
- 184. Nishiyama N, Yokoyama M, Aoyagi T, Okano T, Sakurai Y, Kataoka K. Preparation and characterization of self-assem-

- bled polymer-metal complex micelle from cis-dichlorodiammineplatinum(II) and poly(ethylene glycol)-poly(alpha,beta-aspartic acid) block copolymer in an aqueous medium Langmuir 1999;15:377–83.
- 185. Bontha S, Kabanov AV, Bronich TK. Polymer micelles with cross-linked ionic cores for delivery of anticancer drugs. J Control Release 2006;114(2):163-74.
- 186. Kim JH, Kim YS, Park K, Lee S, Nam HY, Min KH, et al. Antitumor efficacy of cisplatin-loaded glycol chitosan nanoparticles in tumor-bearing mice. J Control Release 2008;127(1):41-9.
- 187. Fang JY, Chen JP, Leu YL, Hu JW. The delivery of platinum drugs from thermosensitive hydrogels containing different ratios of chitosan. Drug Deliv 2008;15(4):235-43.
- 188. H s u SH, Leu YL, Hu JW, Fang JY. Physicochemical characterization and drug release of thermosensitive hydrogels composed of a hyaluronic acid/pluronic f127 graft. Chem Pharm Bull (Tokyo) 2009;57(5):453-8.
- 189. Schechter B, Neumann A, Wilchek M, Arnon R. Soluble polymers as carriers of cis-platinum. J Control Release 1989;10:75–87.
- 190. Bharali DJ, Khalil M, Gurbuz M, Simone TM, Mousa SA. Nanoparticles and cancer therapy: a concise review with emphasis on dendrimers. Int J Nanomedicine 2009;4:1-7.
- 191. Gillies ER, Frechet JM. Dendrimers and dendritic polymers in drug delivery. Drug Discov Today 2005;10(1):35-43.
- 192. Guo R, Shi X. Dendrimers in cancer therapeutics and diagnosis. Curr Drug Metab 2012;In press.
- 193. Gupta U, Dwivedi SK, Bid HK, Konwar R, Jain NK. Ligand anchored dendrimers based nanoconstructs for effective targeting to cancer cells. Int J Pharm 2010;393(1-2):185-96.
- 194. Gul-e-Saba, Abdah A, Abdullah MA. Hyaluronan-mediated CD44 receptor cancer cells progression and the application of controlled drug-delivery system Int J Curr Chem 2010;1(4):195-214.
- 195. Kolhe P, Khandare J, Pillai O, Leih-Lai M, Kannan S, Kannan RM. Hyperbranched polymer–drug conjugates with high drug payload for enhanced cellular delivery. Pharm Res 2004;21:2185–95.

- 196. Perumal O, Khandare J, Kolhe P, Kannan S, Lieh-Lai M, Kannan RM. Effects of branching architecture and linker on the activity of hyperbranched polymer–drug conjugates. Bioconjugate Chem 2009;20:842–6.
- 197. Li X, Qian Y, Liu T, Hu X, Zhang G, You Y, et al. Amphiphilic multiarm star block copolymer-based multifunctional unimolecular micelles for cancer targeted drug delivery and MR imaging. Biomaterials 2011;32(27):6595-605.
- 198. Prabaharan M, Grailer JJ, Pilla S, Steeber DA, Gong S. Amphiphilic multi-arm-block copolymer conjugated with doxorubicin via pH-sensitive hydrazone bond for tumor-targeted drug delivery. Biomaterials 2009;30(29):5757-66.
- 199. Prabaharan M, Grailer JJ, Pilla S, Steeber DA, Gong S. Folate-conjugated amphiphilic hyperbranched block copolymers based on Boltorn H40, poly(L-lactide) and poly(ethylene glycol) for tumor-targeted drug delivery. Biomaterials 2009;30(16):3009-19.
- 200. Tian H, Chen X, Lin H, Deng C, Zhang P, Wei Y, et al. Micellization and reversible pH-sensitive phase transfer of the hyperbranched multiarm PEI-PBLG Copolymer. Chemistry 2006;12(16):4305-12.
- 201. Wang J, del Rosario LS, Demirdirek B, Bae A, Uhrich KE. Comparison of PEG chain length and density on amphiphilic macromolecular nanocarriers: self-assembled and unimolecular micelles. Acta Biomater 2009;5(3):883-92.
- 202. Zou J, Zhao Y, Shi W. Encapsulation mechanism of molecular nanocarriers based on unimolecular micelle forming dendritic coreshell structural polymers. J Phys Chem B 2006;110(6):2638-42.
- 203. Kowalczuk A, Stoyanova E, Mitova V, Shestakova P, Momekov G, Momekova D, et al. Star-shaped nano-conjugates of cisplatin with high drug payload. Int J Pharm 2011;404(1-2):220-30.
- 204. Momekov G, Momekova D, Koseva N, Mitova V, Kowalczuk A, Stoyanova E. Oncopharmacological and biopharmaceutical evaluation of new polymeric delivery systems of platinum-based antineoplastic agents. Polymers 2012 XVII National Symposium Open to International Participation. Ribaritsa, Bulgaria, 2012:22.

- 205. Biros SM, Rebek J, Jr. Structure and binding properties of water-soluble cavitands and capsules. Chem Soc Rev 2007;36(1):93-104.
- 206. Sebestik J, Niederhafner P, Jezek J. Peptide and glycopeptide dendrimers and analogous dendrimeric structures and their biomedical applications. Amino Acids 2011;40(2):301-70.
- 207. Uekama K, Hirayama F, Irie T. Cyclodextrin drug carrier systems. Chem Rev 1998;98(5):2045-76.
- 208. Da Silva E, Lazar AN, Coleman AW. Biopharmaceutical applications of calixarenes. J Drug Deliv Sci Tech 2004;14:3-20.
- 209. Perret F, Coleman AW. Biochemistry of anionic calix[n]arenes. Chem Commun (Camb) 2011;47(26):7303-19.
- 210. Perret F, Lazar AN, Coleman AW. Biochemistry of the para-sulfonato-calix[n]arenes. Chem Commun (Camb) 2006(23):2425-38.
- 211. Casnati A, Sansone F, Ungaro R. Peptidoand glycocalixarenes: playing with hydrogen bonds around hydrophobic cavities. Acc Chem Res 2003;36(4):246-54.
- 212. Casnati A, Sciotto D, Arena G. Water soluble calixarenes. In: Asfari Z, Böhmer V, Harrowfield J, Vicens J, eds. Calixarenes. Dordrecht: Kluwer 2001:440-56.
- 213. Pedersen CJ. The discovery of crown ethers. Science (New York) 1988;241(4865):536-40.
- 214. A n o n y m o u s . Supplementary drugs and other substances: Cyclodextrins. In: Swettman SC, ed. Martindale: The Complete Drug Reference, 36th Ed. London: Pharmaceutical Press 2009:2291.
- 215. R o w e RC, Sheskey PJ, Quinn ME, eds. Handbook of Pharmaceutical Excipients 6th Edition. London. Chicago: Pharmaceutical Press 2009.
- 216. Horvath G, Premkumar T, Boztas A, Lee E, Jon S, Geckeler KE. Supramolecular nanoencapsulation as a tool: solubilization of the anticancer drug trans-dichloro(dipyridine) platinum(II) by complexation with beta-cyclodextrin. Mol Pharm 2008;5(2):358-63.
- 217. Wheate NJ. Improving platinum(II)-based anticancer drug delivery using cucurbit[n]urils. J Inorg Biochem 2008;102(12):2060-6.
- 218. Wheate NJ, Buck DP, Day AI, Collins JG. Cucurbit[n]uril binding of platinum anticancer complexes. Dalton Trans 2006(3):451-8.

- 219. Kennedy AR, Florence AJ, McInnes FJ, Wheate NJ. A chemical preformulation study of a host-guest complex of cucurbit[7]uril and a multinuclear platinum agent for enhanced anticancer drug delivery. Dalton Trans 2009(37):7695-700.
- 220. Jung H, Park KM, Yang JA, Oh EJ, Lee DW, Park K, et al. Theranostic systems assembled in situ on demand by host-guest chemistry. Biomaterials 2011;32(30):7687-94.
- 221. Lee HK, Park KM, Jeon YJ, Kim D, Oh DH, Kim HS, et al. Vesicle formed by amphiphilc cucurbit[6]uril: versatile, noncovalent modification of the vesicle surface, and multivalent binding of sugar-decorated vesicles to lectin. J Am Chem Soc 2005;127(14):5006-7.
- 222. Je o n YJ, Kim SY, Ko YH, Sakamoto S, Yamaguchi K, Kim K. Novel molecular drug carrier: encapsulation of oxaliplatin in cucurbit[7]uril and its effects on stability and reactivity of the drug. Org Biomol Chem 2005;3(11):2122-5.
- 223. Plumb JA, Venugopal B, Oun R, Gomez-Roman N, Kawazoe Y, Venkataramanan NS, et al. Cucurbit[7]uril encapsulated cisplatin overcomes cisplatin resistance via a pharmacokinetic effect. Metallomics 2012;4(6):561-7.
- 224. Wheate NJ, Day AI, Blanch RJ, Arnold AP, Cullinane C, Collins JG. Multi-nuclear platinum complexes encapsulated in cucurbit[n]uril as an approach to reduce toxicity in cancer treatment. Chem Commun (Camb) 2004(12):1424-5.
- 225. Wheate NJ, Taleb RI, Krause-Heuer AM, Cook RL, Wang S, Higgins VJ, et al. Novel platinum(II)-based anticancer complexes and molecular hosts as their drug delivery vehicles. Dalton Trans 2007(43):5055-64.
- 226. Shahgaldian P, Sciotti MA, Pieles U. Amino-substituted amphiphilic calixarenes: self-assembly and interactions with DNA. Langmuir 2008;24(16):8522-6.
- 227. Kunsagi-Mate S, Szabo K, Lemli B, Bitter I, Nagy G, Kollar L. Host-guest interaction between water-soluble calix[6]arene hexasulfonate and p-nitrophenol. Thermochim Acta 2005;425:121-6.
- 228. Martin AD, Raston CL. Multifunctional p-phosphonated calixarenes. Chem Commun (Camb) 2011;47(35):9764-72.
- 229. Ukhatskaya EV, Kurkov SV, Matthews SE, El Fagui A, Amiel C, Dalmas F, et al. Eval-

- uation of a cationic calix[4]arene: Solubilization and self-aggregation ability. Int J Pharm 2010;402(1-2):10-9.
- 230. Dudic M, Colombo A, Sansone F, Casnati A, Donofrio G, Ungaro R. A general synthesis of water soluble upper rim calix[n]arene guanidinium derivatives which bind to plasmid DNA. Tetrahedron 2004;60:11613-8
- 231. Sansone F, Chierici E, Casnati A, Ungaro R. Thiourea-linked upper rim calix[4]arene neoglycoconjugates: synthesis, conformations and binding properties. Org Biomol Chem 2003;1(10):1802-9.
- 232. Fulton DA, Stoddart JF. Neoglycoconjugates based on cyclodextrins and calixarenes. Bioconjug Chem 2001;12(5):655-72.
- 233. Taton D, Saule M, Logan J, Duran R, Hou S, Chaikof EL, et al. Polymerization of ethylene oxide with a calixarene-based precursor: Synthesis of eight-arm poly(ethylene oxide) stars by the core-first methodology. J Polymer Sci Part A: Polymer Chem 2003;41(21):1669–76.

- 234. Tu C, Zhu L, Li P, Chen Y, Su Y, Yan D, et al. Supramolecular polymeric micelles by the host-guest interaction of star-like calix[4] arene and chlorin e6 for photodynamic therapy. Chem Commun (Camb) 2010;47(21):6063-5.
- 235. Momekov G, Budurova D, Drakalska E, Shenkov S, Momekov G, Trzebicka B, et al. Aggregation behavior and in vitro biocompatibility study of octopus-shaped macromolecules based on tert-butylcalix[4]arenes. Int J Pharm 2012;436(1-2):410-7.
- 236. Krause-Heuer AM, Wheate NJ, Tilby MJ, Pearson DG, Ottley CJ, Aldrich-Wright JR. Substituted beta-cyclodextrin and calix[4]arene as encapsulatory vehicles for platinum(II)-based DNA intercalators. Inorg Chem 2008;47(15):6880-8.
- 237. Ding J, Pan D, Tung CH, Wu LZ. Synthesis and photophysical studies of calix[4]arenebased binuclear platinum(II) complexes: probing metal-metal and ligand-ligand interactions. Inorg Chem 2008;47(12):5099-106.

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